

Ecology of *Salmonella*

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Salmonella evolution

- Evolved over the past 12—160 million years! (split between *E. coli* and *Salmonella*)
- Can infect >100 different species including mammals, birds, reptiles, and insects
- Complex nomenclature:
 - Two major species: *S. enterica* and *S. bongori*
 - Seven subspecies, including *enterica* (I) accounting for >99% of human infection

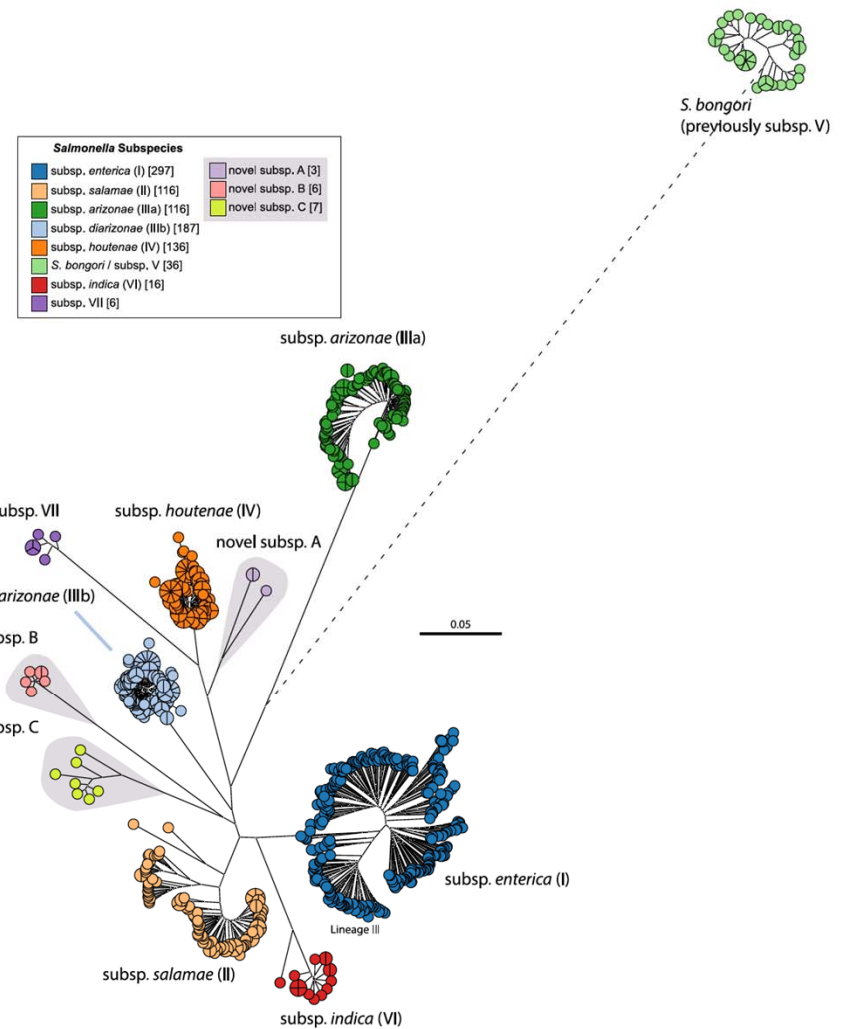
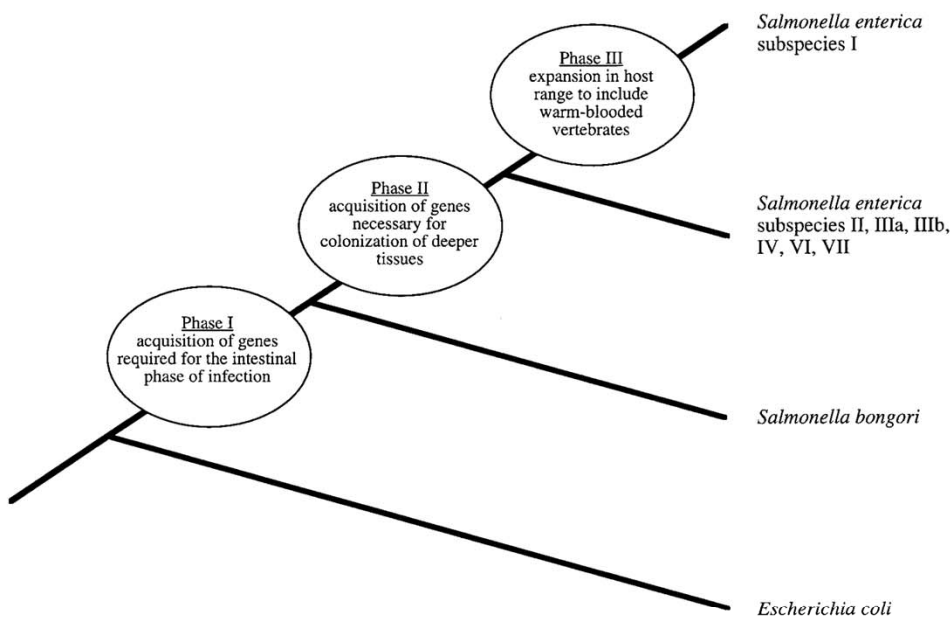
Salmonella evolution

REVIEW

A genomic overview of the population structure of *Salmonella*

Nabil-Fareed Alikhan, Zhemín Zhou, Martin J. Sergeant, Mark Achtman*

Warwick Medical School, University of Warwick, Coventry, United Kingdom

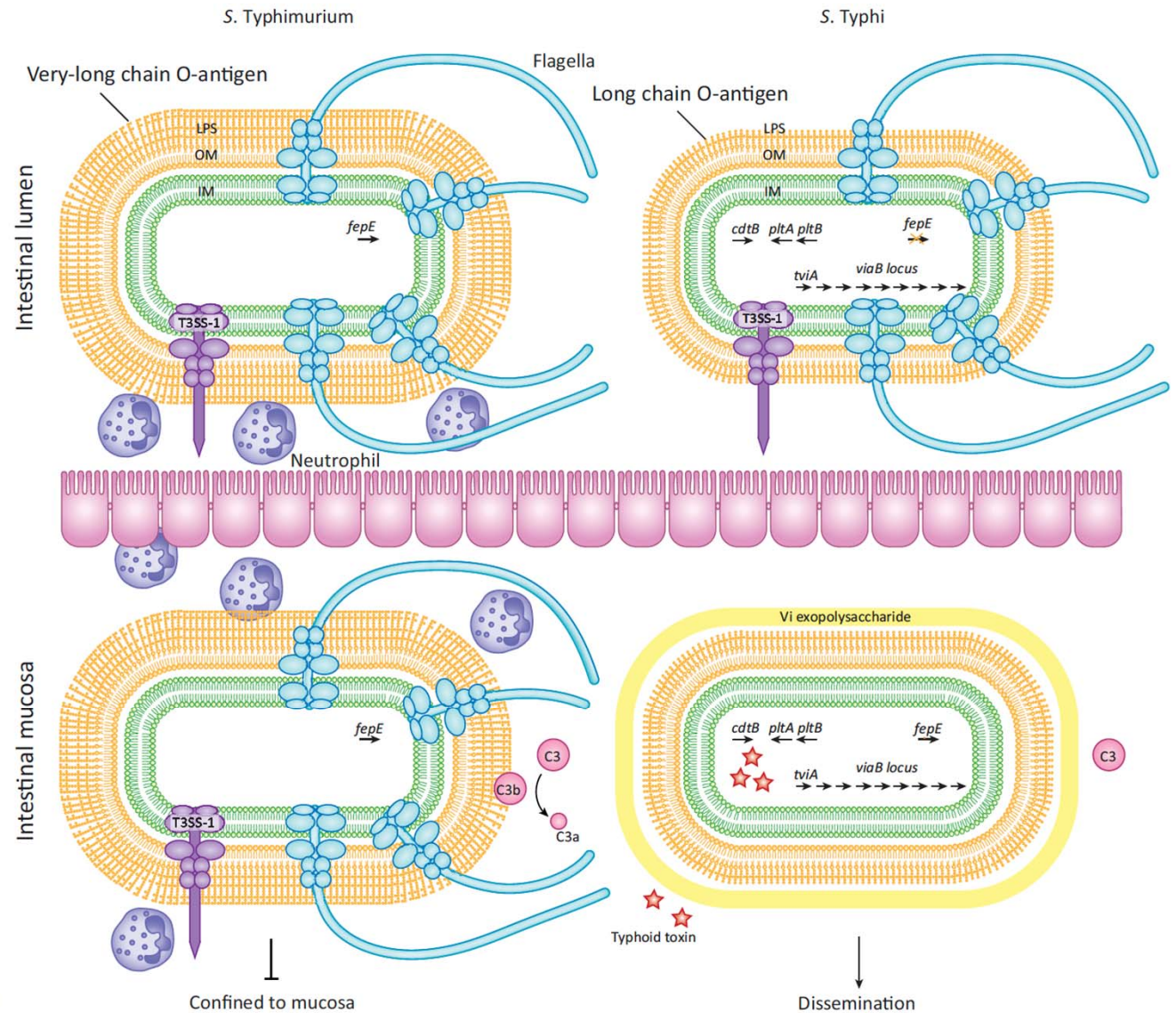


MINIREVIEW

Evolution of Host Adaptation in *Salmonella enterica*

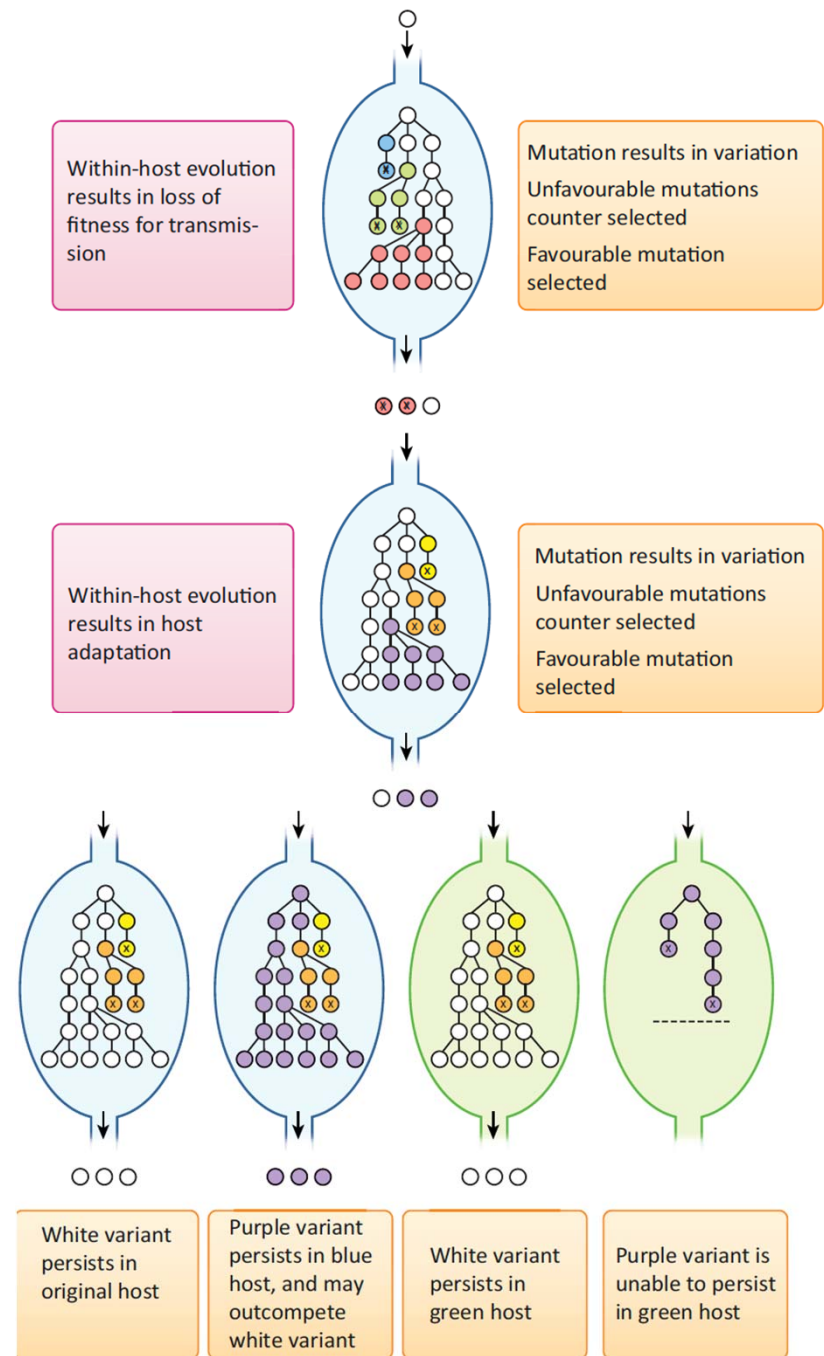
ANDREAS J. BÄUMLER,^{1*} RENÉE M. TSOLIS,² THOMAS A. FICHT,² AND L. GARRY ADAMS²
 Department of Medical Microbiology and Immunology, College of Medicine,¹ and Department of Veterinary Pathobiology, College of Veterinary Medicine,² Texas A&M University, College Station, Texas 77843-4467

Salmonella evolution



Review
Evolution of *Salmonella* within Hosts

Tradeoffs for host range versus host adaptation

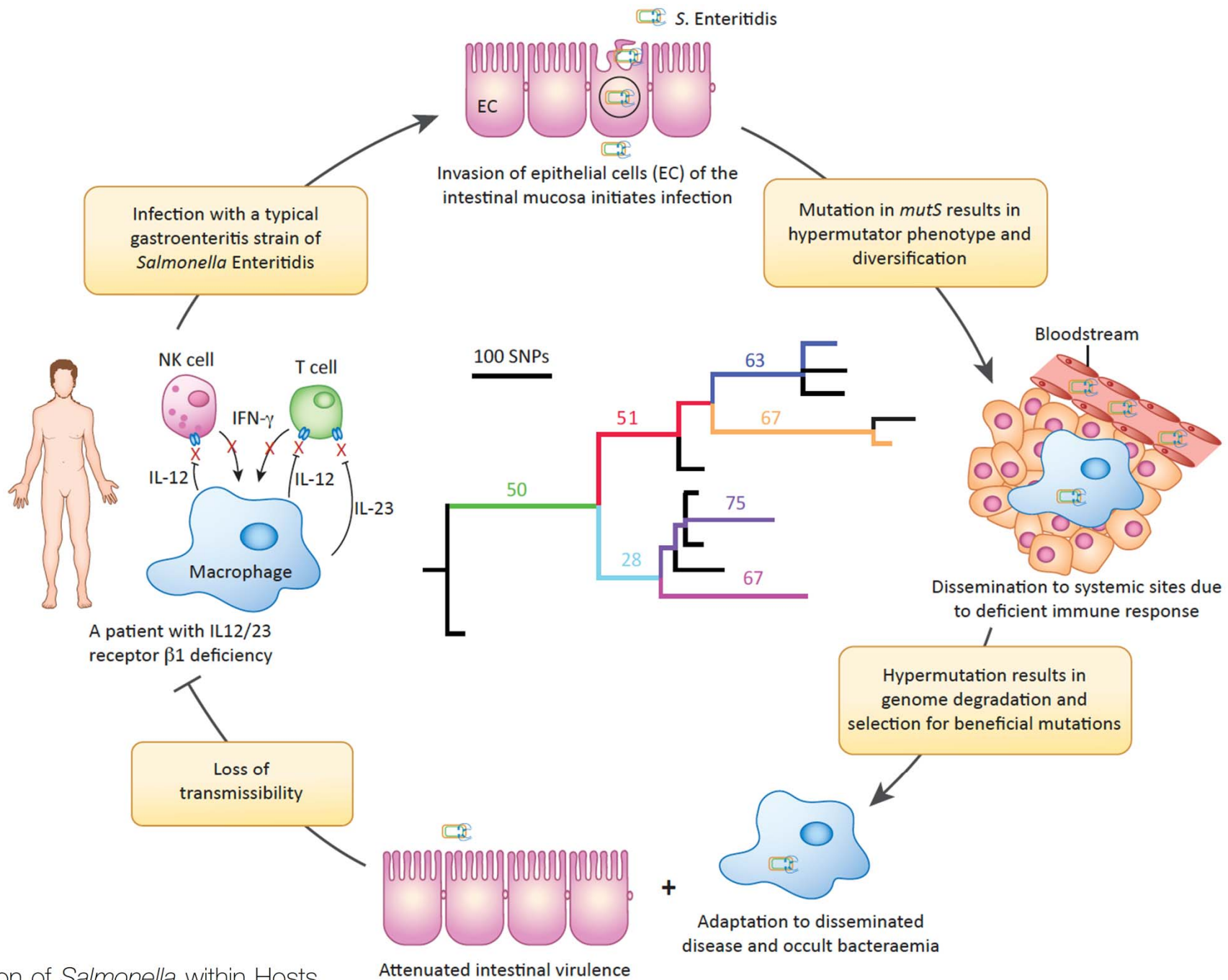


Review

Evolution of *Salmonella* within Hosts

Jennifer R. Tanner¹ and Robert A. Kingsley^{1,*}

Trends in Microbiology



Review
Evolution of *Salmonella* within Hosts

Salmonella typing

- Kauffmann-White serotyping scheme give the serovars their names
 - LPS (O) antigen
 - H1 and H2 flagellar antigens
 - Capsular antigen
- O antigen divides into 6 serogroups: A, B, 1, C2, D, E
- >2,300 serovars

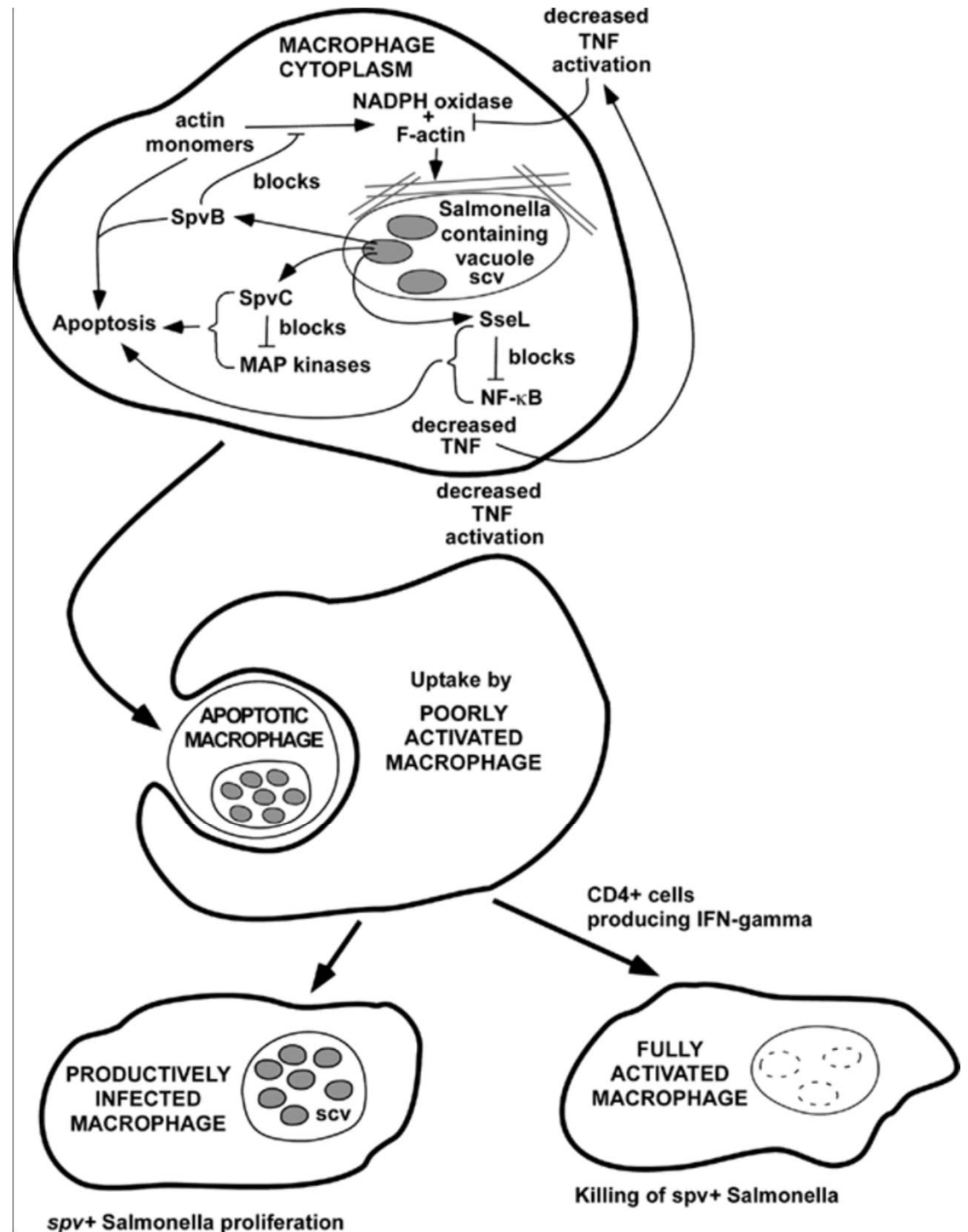
Factors involved in host range

- Colonization
 - SPI-3, SPI-4
 - Numerous fimbrial operons (*agf*, *sef*, *pil*, *lpf*, *pef*)
- Persistence
 - LPS
- Invasion
 - Exotoxins and enterotoxins
 - SPI-1, SPI-5
- Intracellular survival
 - SPI-2 through SPI-6
 - Virulence plasmid (*spv*)
- Phase variation in flagella expression

Plasmids

- Often ignored in outbreak investigations
- They provide critical hints about host source and host range
- They can encode for multidrug resistance
- They can encode for enhanced virulence
- They can enable enhanced fitness

Plasmids – virulence plasmid

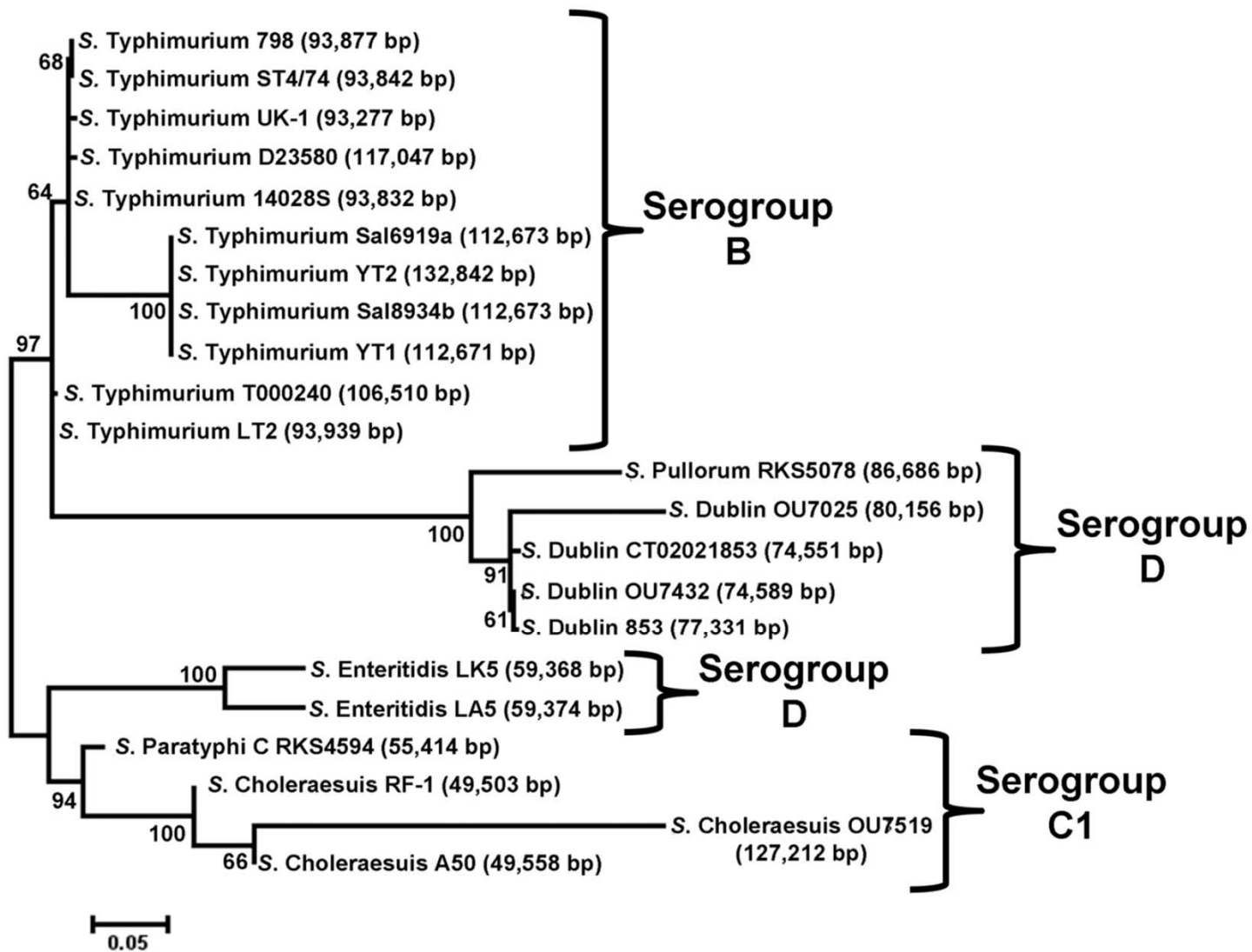


The role of the *spv* genes in *Salmonella* pathogenesis

Donald G. Guiney^{1*} and Joshua Fierer^{1,2}

¹ Department of Medicine, University of California San Diego School of Medicine, La Jolla, CA, USA

² Veterans Administration Medical Center, La Jolla, CA, USA

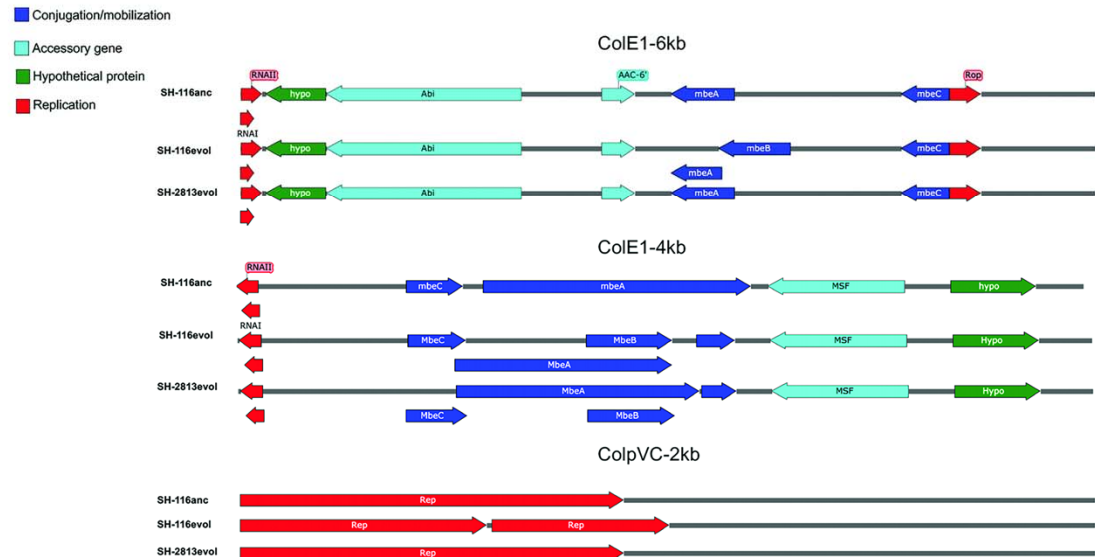
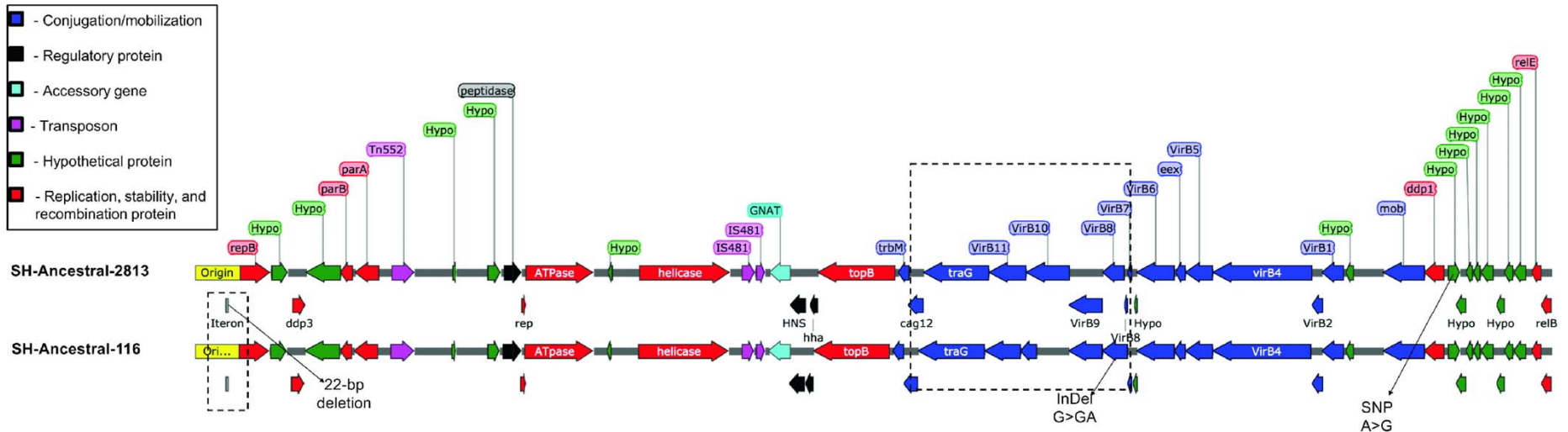


Salmonella Pathogenicity and Host Adaptation in Chicken-Associated Serovars

Steven L. Foley,^a Timothy J. Johnson,^b Steven C. Ricke,^c Rajesh Nayak,^a Jessica Danzeisen^b

Division of Microbiology, National Center for Toxicological Research, U.S. Food and Drug Administration, Jefferson, Arkansas, USA^a; Department of Veterinary and Biomedical Sciences, University of Minnesota, Saint Paul, Minnesota, USA^b; Center for Food Safety and Department of Food Science, University of Arkansas, Fayetteville, Arkansas, USA^c

Plasmids – IncX and Col plasmids

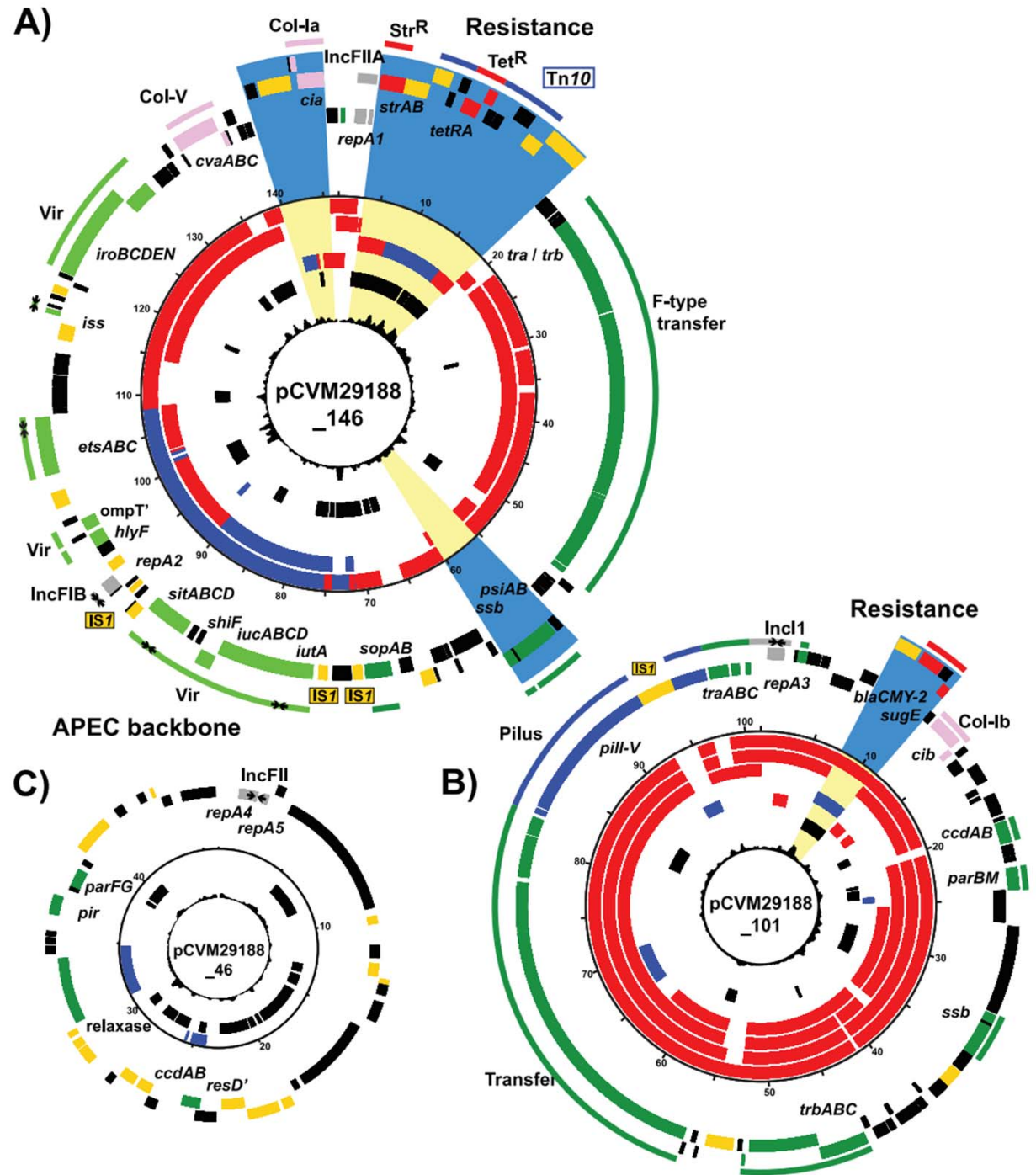


RESEARCH ARTICLE

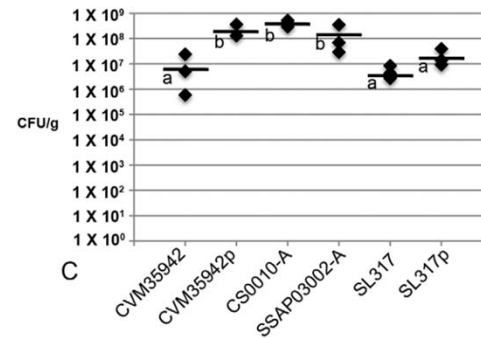
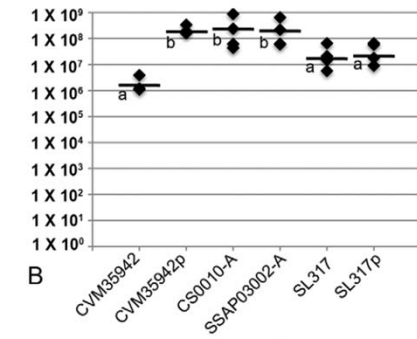
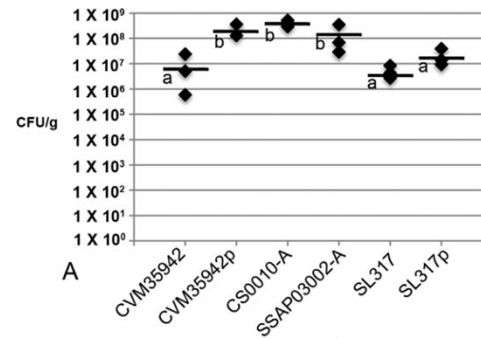
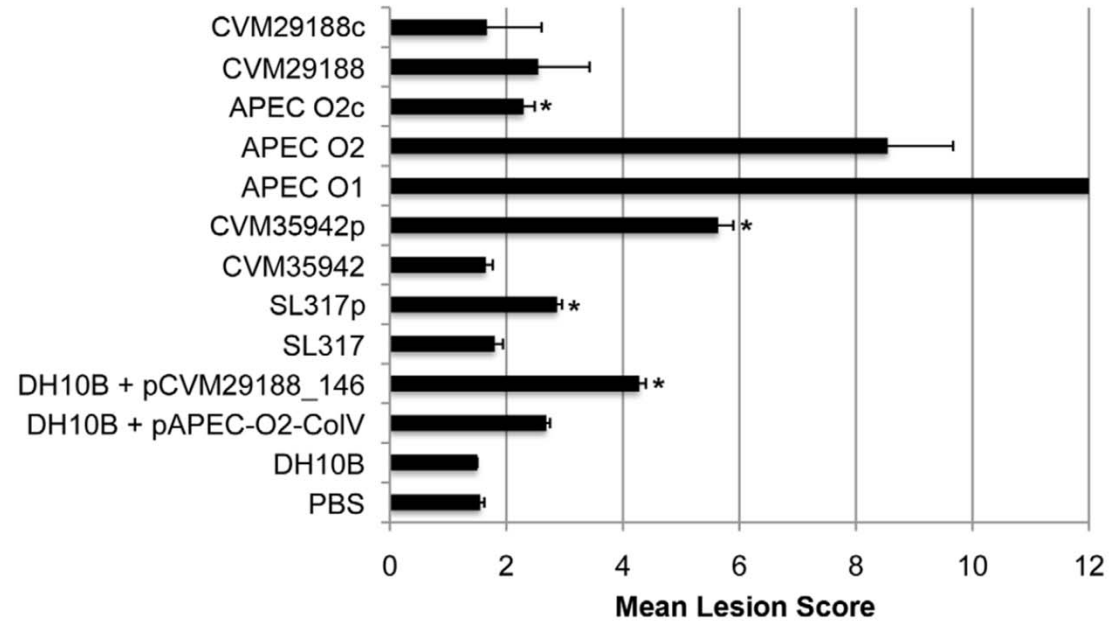
Hotspot mutations and ColE1 plasmids contribute to the fitness of *Salmonella* Heidelberg in poultry litter

Adelumola Oladeinde¹*, Kimberly Cook¹, Alex Orlek², Greg Zock¹,
 Kyler Herrington³, Nelson Cox⁴, Jodie Plumlee Lawrence¹, Carolina Hall¹

Plasmids – ColV plasmid



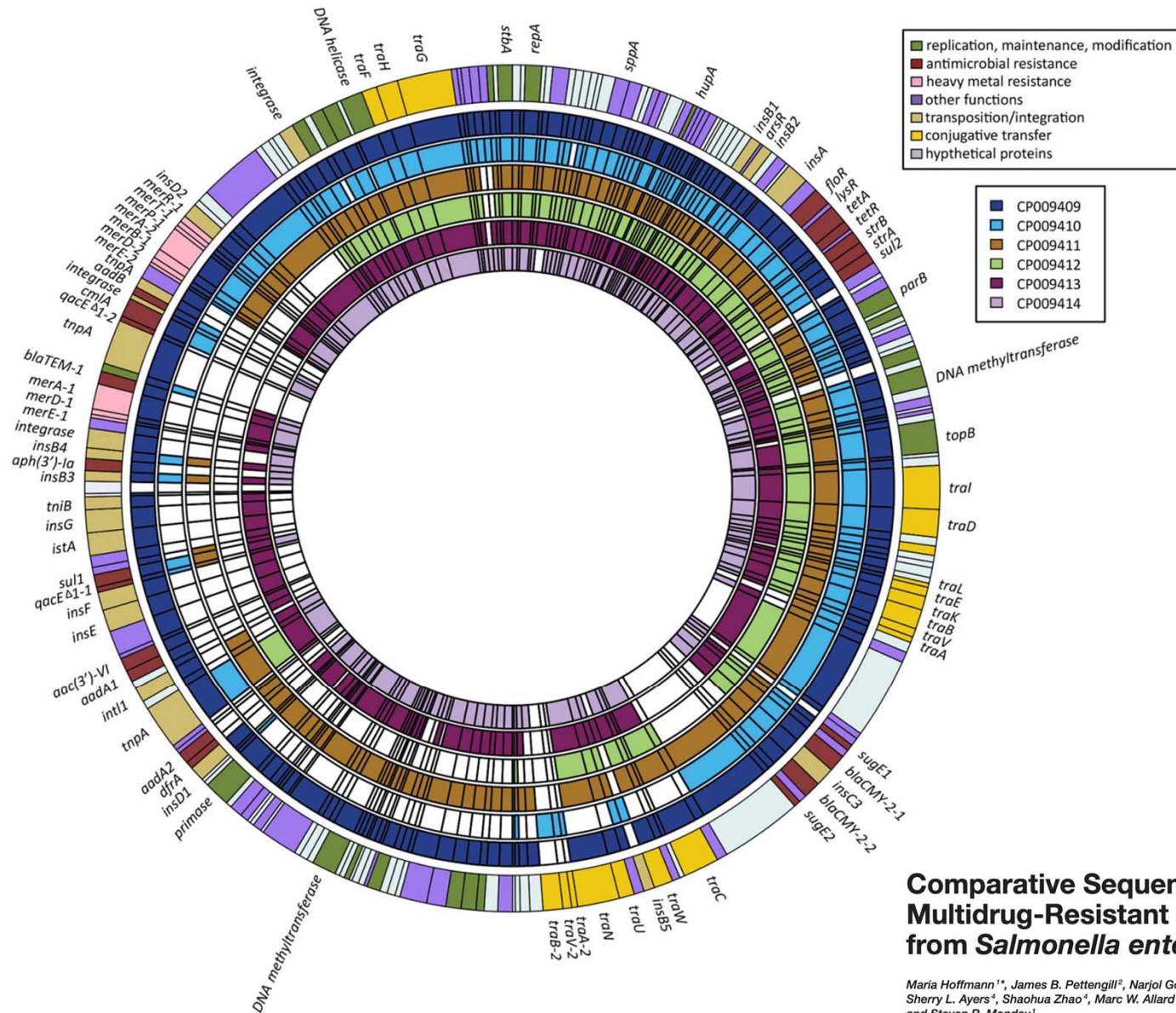
Plasmids – ColV plasmid



Horizontal Gene Transfer of a ColV Plasmid Has Resulted in a Dominant Avian Clonal Type of *Salmonella enterica* Serovar Kentucky

Timothy J. Johnson^{1*}, Jessica L. Thorsness¹, Cole P. Anderson², Aaron M. Lynne², Steven L. Foley³, Jing Han³, W. Florian Fricke⁴, Patrick F. McDermott⁵, David G. White⁵, Mahesh Khatri¹, Adam L. Stell¹, Cristian Flores¹, Randall S. Singer¹

Plasmids – IncA/C plasmid



Comparative Sequence Analysis of Multidrug-Resistant IncA/C Plasmids from *Salmonella enterica*

Maria Hoffmann^{1*}, James B. Pettengill², Narjol Gonzalez-Escalona¹, John Miller^{2,3}, Sherry L. Ayers⁴, Shaohua Zhao⁴, Marc W. Allard¹, Patrick F. McDermott⁴, Eric W. Brown¹ and Steven R. Monday¹

Plasmids – IncA/C plasmid

TABLE 2 | List of the resistance genotype-phenotype identified for the six IncA/C plasmids.

| | S. Kentucky CFSAN007428 | S. Newport CFSAN007425 | S. Typhimurium CFSAN007405 | S. Agona CFSAN007426 | S. Heidelberg CFSAN000405 | S. Infantis CFSAN007427 |
|-------------------------------|---------------------------------------------------------------------------------------------------------------------------|------------------------------------------------------------------------------------------------------------------------|------------------------------------------------------------------------------------------------------------------------|---------------------------------------------------|---------------------------------------------------------------------------------------------------------------------------|---------------------------------------------------------------------------------------------------------------------------|
| Resistance Phenotype | AMC, AMP, FOX, AXO, CHL, GEN, KAN, STR, FIS, TET, TIO | AMC, AMP, FOX, AXO, CEP, CHL, STR, FIS, TET, COT, TIO | AMC, AMP, FOX, AXO, CEP, CHL, KAN, NAL, STR, FIS, TET, TIO | AMC, AMP, FOX, AXO, CHL, STR, FIS, TET, TIO | AMC, AMP, FOX, AXO, CHL, GEN, KAN, STR, FIS, TET, TIO | AMC, AMP, FOX, AXO, CHL, GEN, KAN, STR, FIS, TET, TIO |
| Beta-Lactams | <i>bla_{cmy-2}</i> , <i>bla_{tem-1}</i> | <i>bla_{cmy-2}*</i> | <i>bla_{cmy-2}*</i> | <i>bla_{cmy-2}</i> | <i>bla_{cmy-2}*</i> , <i>bla_{tem-1}</i> | <i>bla_{cmy-2}</i> , <i>bla_{tem-1}</i> |
| Aminoglycosides | <i>strA</i> , <i>strB</i> , <i>aadB</i> <i>aph(3)-Ia</i> | <i>strA</i> , <i>strB</i> , <i>aadA2</i> | <i>strA</i> , <i>strB</i> , <i>aadB</i> | <i>strA</i> , <i>strB</i> | <i>strA</i> , <i>strB</i> , <i>aadA1</i> , <i>aadB</i> , <i>aac(3')-VI</i> , <i>aph(3')-Ia</i> | <i>strA</i> , <i>strB</i> , <i>aadA1</i> , <i>aac(3)-VI</i> |
| Tetracyclines | <i>tetR(A)</i> , <i>tetA</i> | <i>tetR(A)</i> , <i>tetA</i> | <i>tetR(A)</i> , <i>tetA</i> | <i>tetR(A)</i> , <i>tetA</i> | <i>tetR(A)</i> , <i>tetA</i> | <i>tetR(A)</i> , <i>tetA</i> |
| Chloramphenicols | <i>floR</i> , <i>cmlA</i> | <i>floR</i> | <i>floR</i> , <i>cmlA</i> | <i>floR</i> | <i>floR</i> , <i>cmlA</i> | <i>floR</i> |
| Folate synthesis inhibitors | <i>sul1</i> , <i>sul2</i> | <i>sul1</i> , <i>sul2</i> , <i>dfra12</i> | <i>sul1</i> , <i>sul2</i> | <i>sul2</i> | <i>sul1</i> , <i>sul2</i> | <i>sul1</i> , <i>sul2</i> |
| Quaternary ammonium compounds | <i>quacE</i> , <i>sugE</i> | <i>quacE</i> , <i>sugE*</i> | <i>quacE</i> , <i>sugE*</i> | <i>sugE</i> | <i>quacE*</i> , <i>sugE*</i> | <i>quacE</i> , <i>sugE</i> |
| Mercury ions | <i>merA*</i> , <i>merB</i> , <i>merC</i> , <i>merD*</i> , <i>merE*</i> , <i>merP</i> , <i>merT</i> , <i>merR</i> | <i>merA</i> , <i>merB</i> , <i>merC</i> , <i>merD</i> , <i>merE</i> , <i>merP</i> , <i>merT</i> , <i>merR</i> | <i>merA</i> , <i>merB</i> , <i>merC</i> , <i>merD</i> , <i>merE</i> , <i>merP</i> , <i>merT</i> , <i>merR</i> | | <i>merA*</i> , <i>merB</i> , <i>merC</i> , <i>merD*</i> , <i>merE*</i> , <i>merP</i> , <i>merT</i> , <i>merR</i> | <i>merA*</i> , <i>merB</i> , <i>merC</i> , <i>merD*</i> , <i>merE*</i> , <i>merP</i> , <i>merT</i> , <i>merR</i> |

Comparative Sequence Analysis of Multidrug-Resistant IncA/C Plasmids from *Salmonella enterica*

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Restricted versus non-restricted host range

- Those dominating human infection tend to have broad host range
 - Typhimurium, Enteritidis, Newport, Heidelberg
- Others have restricted host range
 - Typhi, Paratyphi, Gallinarum, Pullorum, Choleraesuis, Dublin
- When spillover of restricted serovars occurs, they tend to be more invasive
- Most of the host-adapted serovars have tendency to accumulate pseudogenes

| Host species | Disease | <i>S. enterica</i> subspecies I serotype(s) most frequently encountered | Most susceptible age groups | Typical symptoms or sign(s) of disease | Reference |
|--------------|----------------------|-------------------------------------------------------------------------|-----------------------------|----------------------------------------------------|-----------|
| Humans | Salmonella enteritis | Typhimurium, Enteritidis | Children (<4 yr) | Diarrhea, dysentery, fever | 78 |
| | Typhoid fever | Typhi ^c | Children and adults | Septicemia, fever ^a | 78 |
| | Paratyphoid fever | Sendai; Paratyphi A, B, and C ^c | Children and adults | Septicemia, fever ^a | 78 |
| Cattle | Salmonellosis | Typhimurium | Calves (<8 wk) | Diarrhea, dysentery, septicemia, fever | 104 |
| | | Dublin | Calves and adult cattle | Diarrhea, dysentery, septicemia, abortion, fever | 99, 104 |
| Poultry | Pullorum disease | Pullorum ^{c,d} | Newly hatched birds | Diarrhea, septicemia | 27 |
| | Fowl typhoid | Gallinarum ^{c,d} | Growing stock and adults | Diarrhea, comb discoloration, septicemia | 27 |
| | Avian paratyphoid | Enteritidis, Typhimurium | Newly hatched birds | Diarrhea, septicemia | 27 |
| Sheep | Salmonellosis | Abortusovis ^c | Adult sheep | Septicemia, abortion, vaginal discharge | 90 |
| | | Typhimurium | Lambs | Diarrhea, dysentery, septicemia | 90 |
| | | | Lambs | Diarrhea, dysentery, septicemia | 90 |
| Pigs | Pig paratyphoid | Choleraesuis ^c | Weaned and adult pigs | Skin discoloration, septicemia, fever ^b | 106 |
| | Salmonellosis | Typhimurium | Weaned pigs (<4 mo) | Diarrhea | 106 |
| | Chronic paratyphoid | Typhisuis | | Intermittent diarrhea | 7 |
| Horses | Salmonellosis | Abortusequi ^c | Adult horses | Septicemia, abortion | 110 |
| | | | Foals | Diarrhea, septicemia | 110 |
| | | Typhimurium | Foals | Diarrhea, septicemia | 110 |
| Wild rodents | Murine typhoid | Typhimurium, Enteritidis | | Septicemia, fever | 28 |

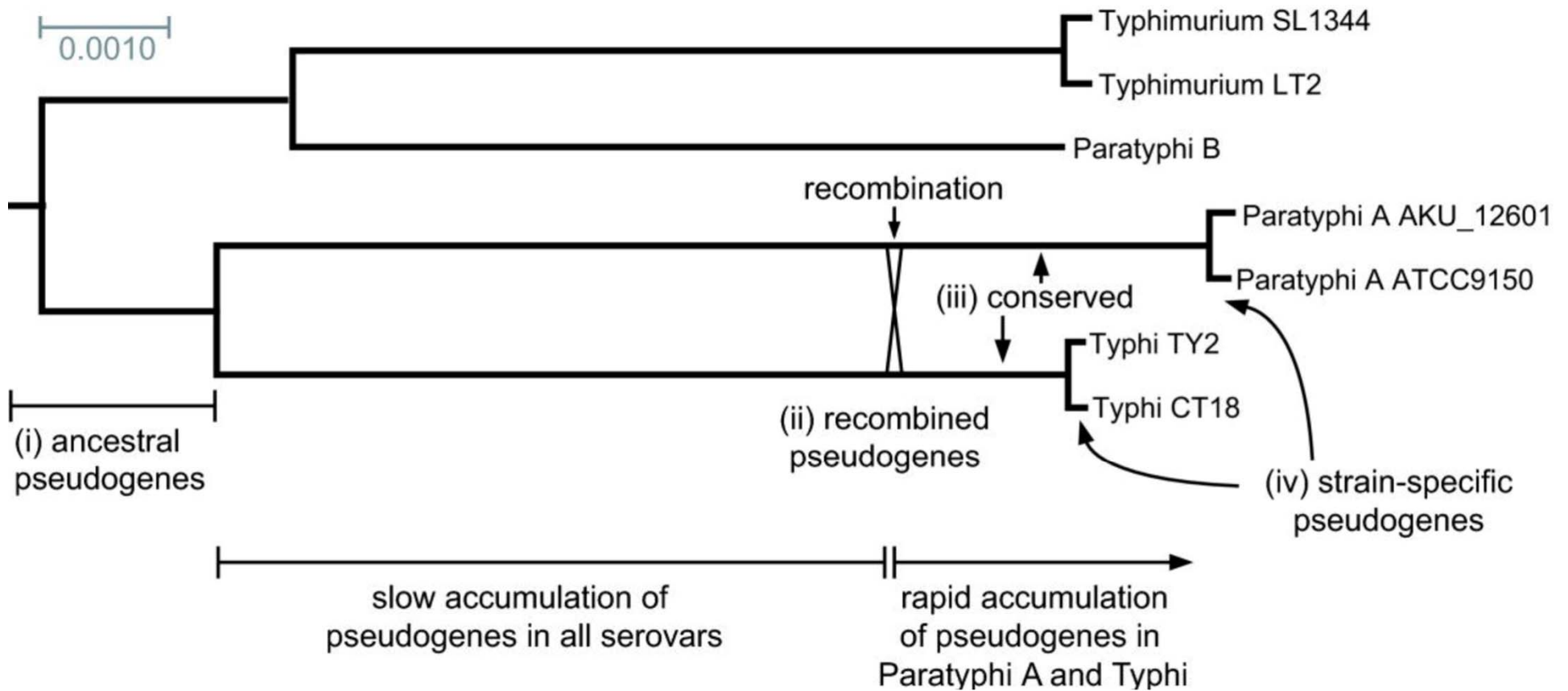
MINIREVIEW

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Department of Medical Microbiology and Immunology, College of Medicine,¹ and Department of Veterinary Pathobiology, College of Veterinary Medicine,² Texas A&M University, College Station, Texas 77843-4467

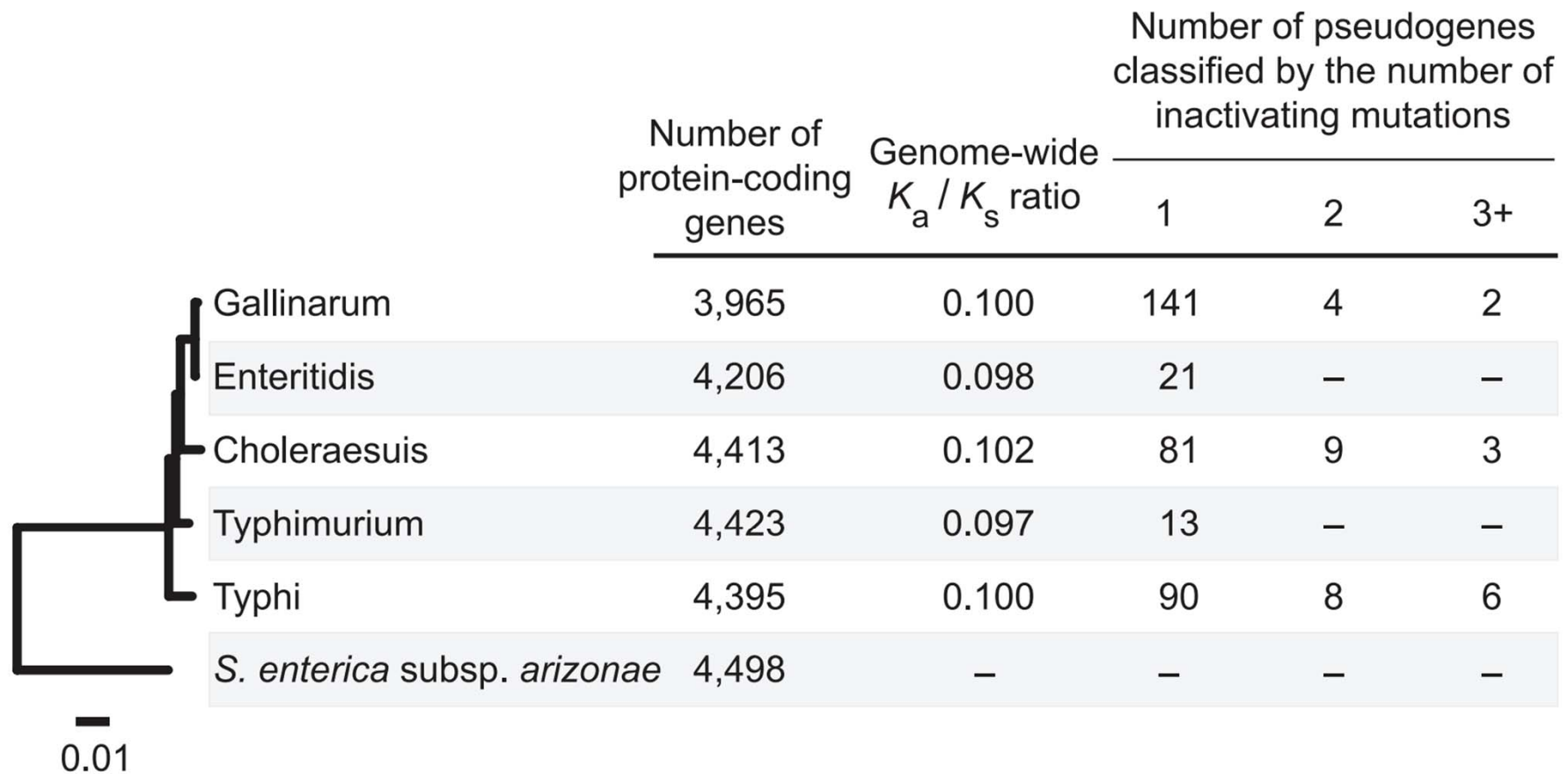
Pseudogene formation in *Salmonella*



Pseudogene accumulation in the evolutionary histories of *Salmonella enterica* serovars Paratyphi A and Typhi

Kathryn E Holt, Nicholas R Thomson, John Wain, Gemma C Langridge, Rumina Hasan, Zulfiqar A Bhutta, Michael A Quail, Halina Norbertczak, Danielle Walker, Mark Simmonds, Brian White, Nathalie Bason, Karen Mungall, Gordon Dougan and Julian Parkhill

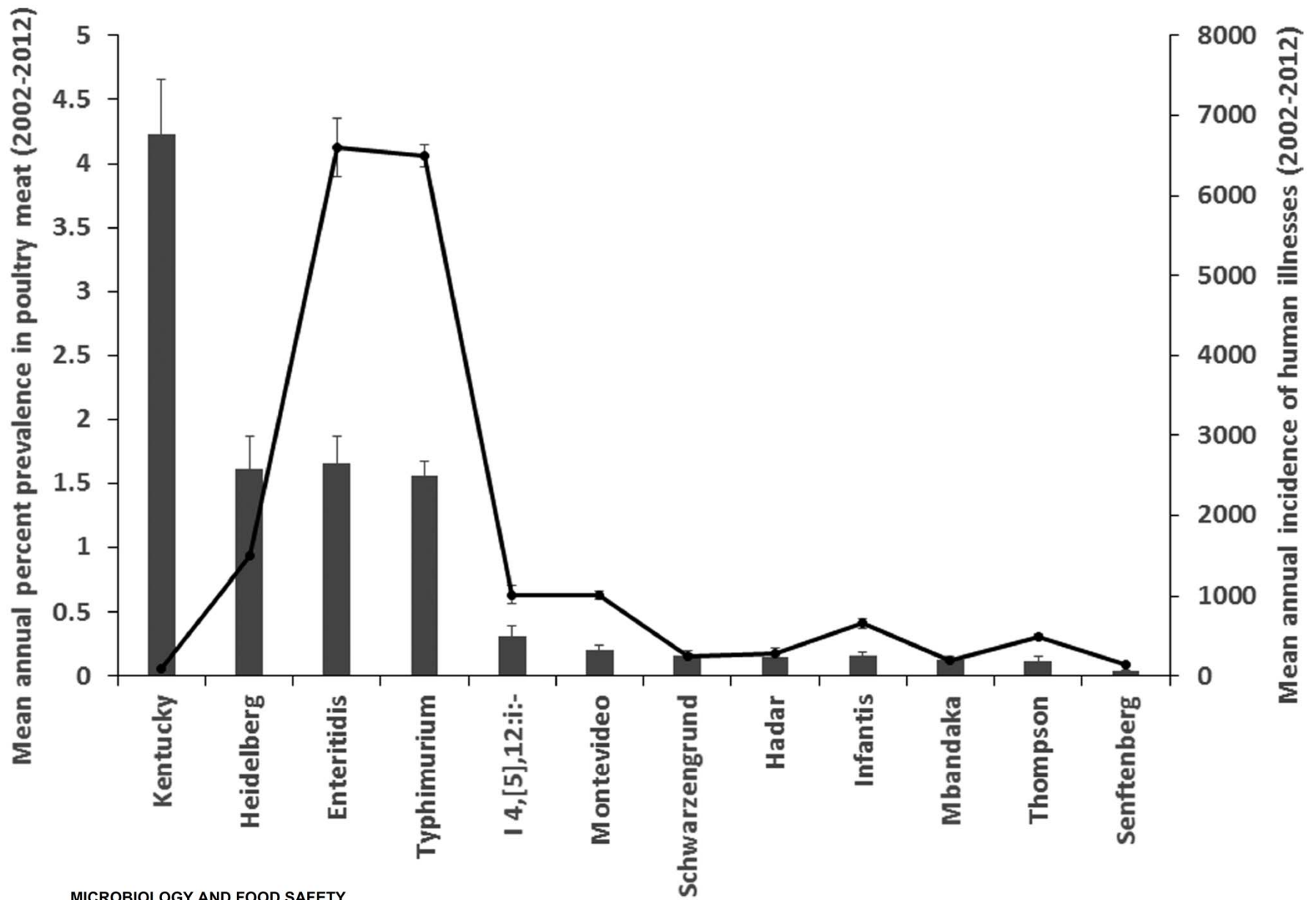
Pseudogene formation in *Salmonella*



The Extinction Dynamics of Bacterial Pseudogenes

Chih-Horng Kuo¹, Howard Ochman*

¹Department of Ecology and Evolutionary Biology, University of Arizona, Tucson, Arizona, United States of America

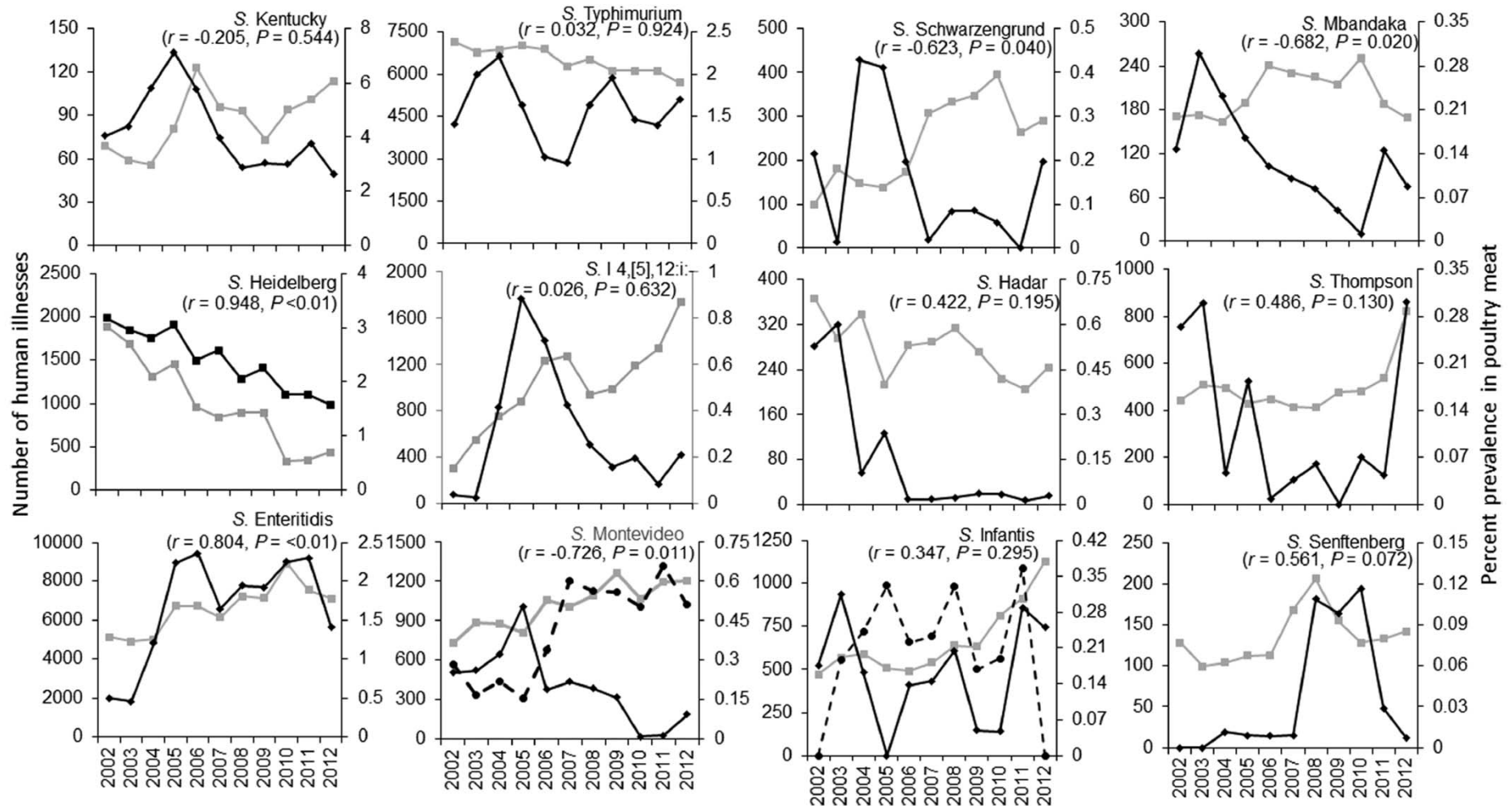


MICROBIOLOGY AND FOOD SAFETY

Population dynamics and antimicrobial resistance of the most prevalent poultry-associated *Salmonella* serotypes

Devendra H. Shah,^{*1} Narayan C. Paul,^{*} William C. Sicho,¹ Rocio Crespo,^{*} and Jean Guard¹

^{*}Department of Veterinary Microbiology and Pathology; ¹Department of Veterinary Clinical Sciences, Washington State University, Pullman, WA 99164-7040; and ³Egg Quality and Safety Research Unit, United States Department of Agriculture, Atlanta, GA 30605, USA



MICROBIOLOGY AND FOOD SAFETY

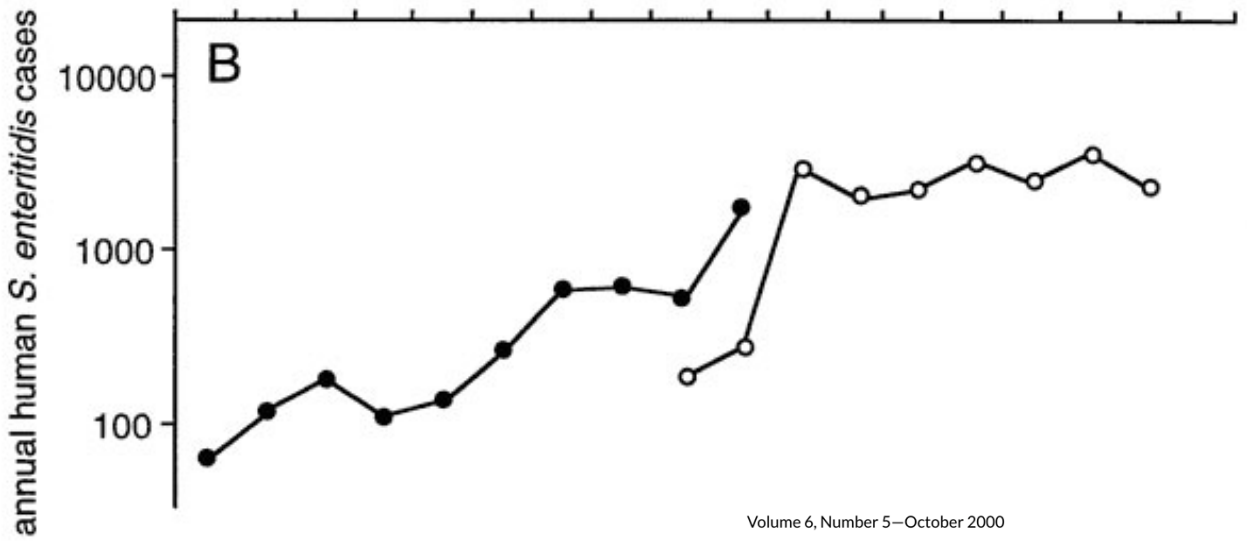
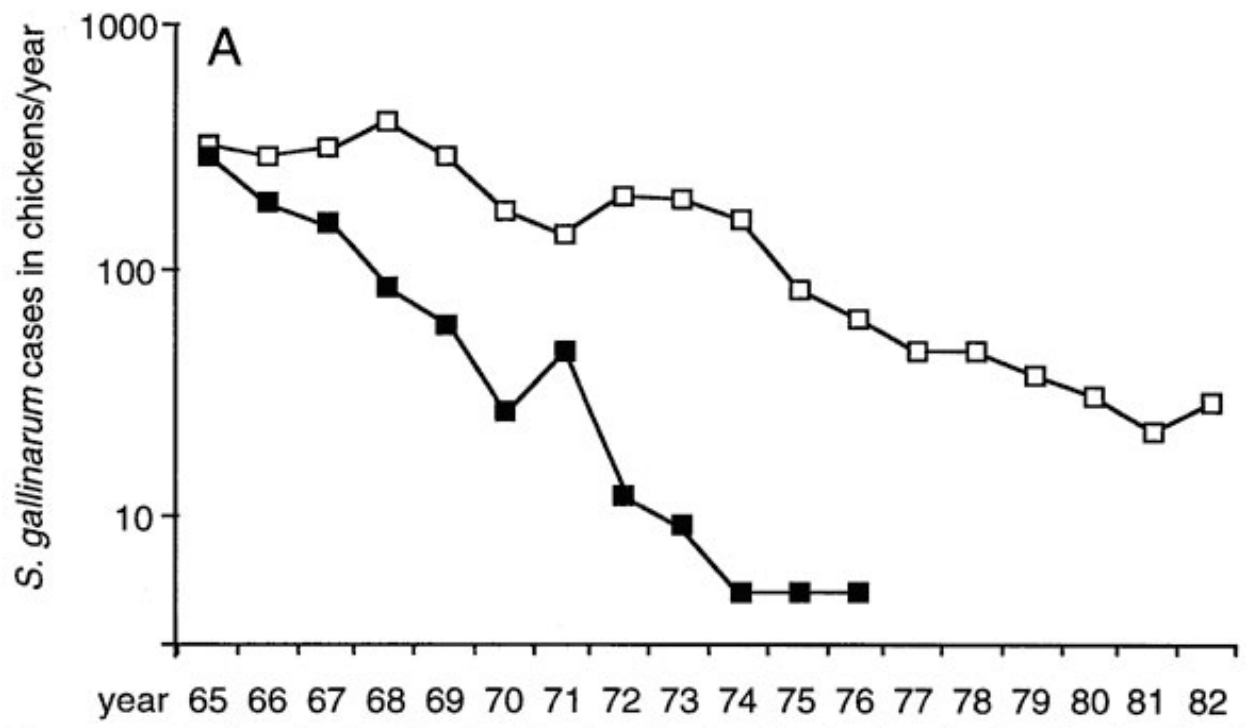
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Niche replacement

- Principle of competitive exclusion applies to Salmonella
- Example: *Salmonella* Gallinarum, Pullorum, and Enteritidis
 - All are O9 antigen
 - Gallinarum and Pullorum were deadly to poultry and were eradicated
 - Enteritidis now occupies that niche
 - Enteritidis has tropism for the reproductive tract of the chicken
 - Spillover into table eggs occurs more frequently than pre-eradication of Gallinarum/Pullorum



Volume 6, Number 5—October 2000

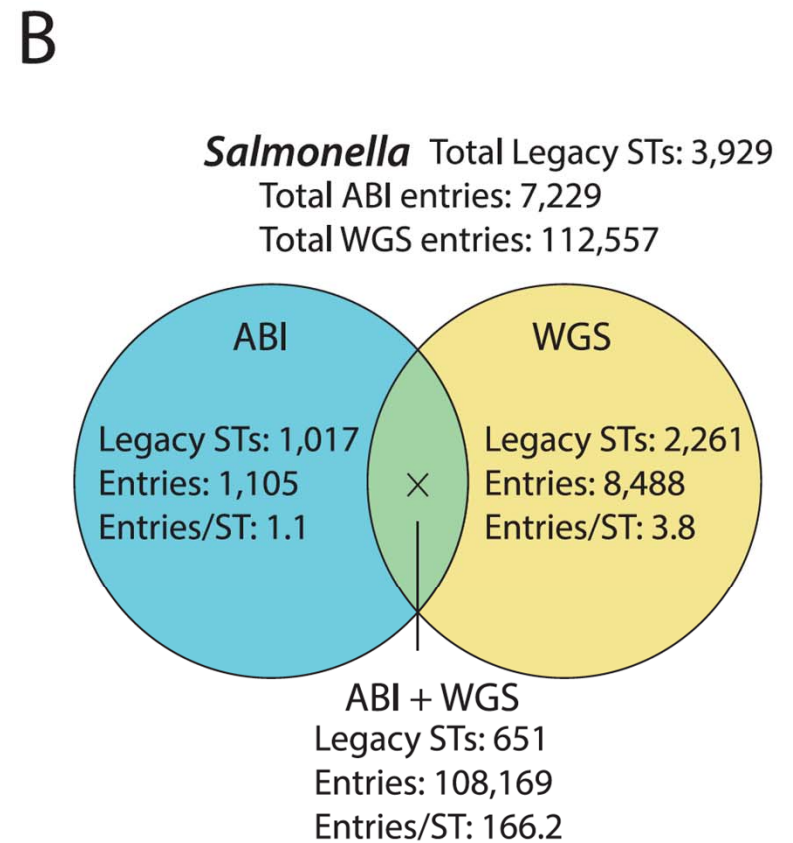
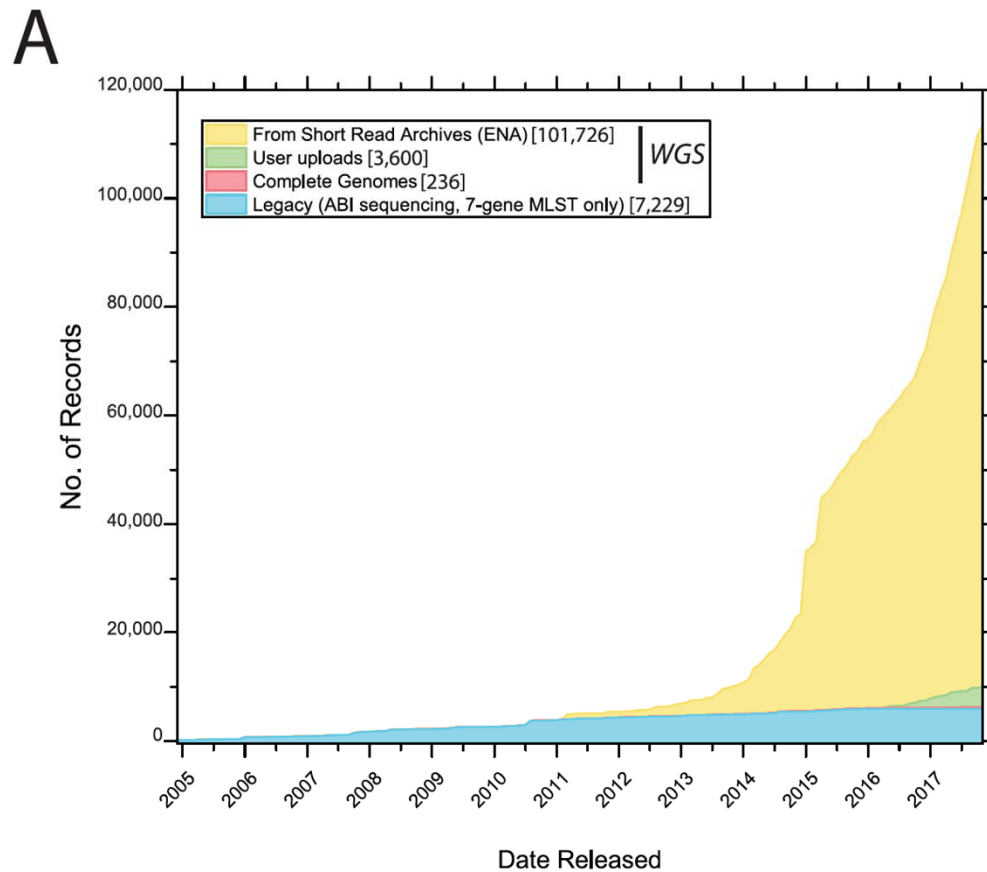
Perspective

Competitive Exclusion of *Salmonella* Enteritidis by *Salmonella* Gallinarum in Poultry

Wolfgang Rabsch¹, Billy M. Hargis², Renée M. Tsolis¹, Robert A. Kingsley¹, Karl-Heinz Hinz³, Helmut Tschäpe⁴, and Andreas J. Baumler¹□

Author affiliations: ¹Robert Koch Institute, Wernigerode, Germany; ²Texas A&M University, College Station, Texas, USA; ³School of Veterinary Medicine, Hanover, Germany

Whole genome sequencing is here!



REVIEW

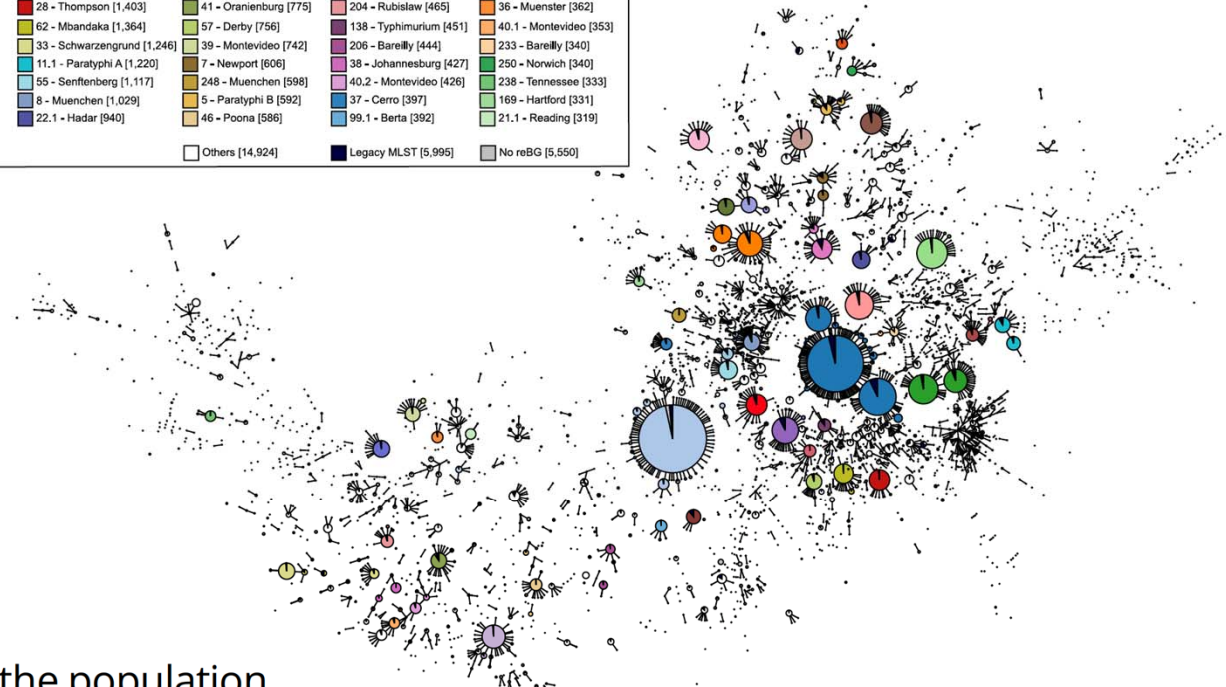
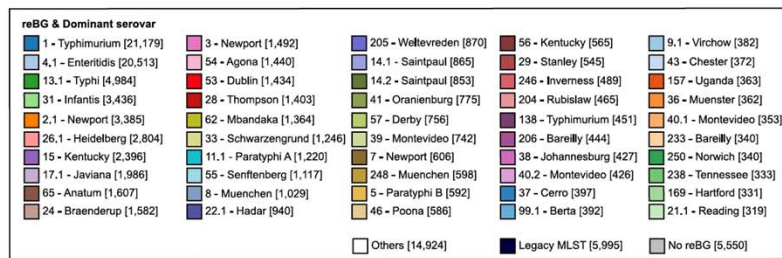
A genomic overview of the population structure of *Salmonella*

Nabil-Fareed Alikhan, Zheming Zhou, Martin J. Sergeant, Mark Achtman*

Warwick Medical School, University of Warwick, Coventry, United Kingdom

Table 1. *Salmonella* MLST genotyping schemes offered by EnteroBase (1 November, 2017).

| Legacy MLST | rMLST | cgMLST | wgMLST |
|-----------------------------------|------------------------------------|-----------------------------------------|----------------------------------------------------------------------------|
| 7 Loci | 51 Loci | 3,002 Loci | 21,065 Loci |
| Conserved housekeeping genes | Ribosomal genes | A soft subset of core genes from wgMLST | All pan-genomic coding sequences from 537 completed/representative genomes |
| Conserved low resolution | Highly conserved medium resolution | Variable high resolution | Ultravariabile extreme resolution |
| Different scheme for each species | Single scheme across tree of life | Different scheme for each genus | Different scheme for each genus |
| STs: 3,929 eBGs: 360 | rSTs: 5,454 reBGs: 337 | cgSTs: 96,108 | wgSTs: 112,409 |



REVIEW

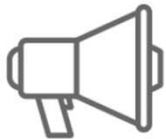
A genomic overview of the population structure of *Salmonella*

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Warwick Medical School, University of Warwick, Coventry, United Kingdom

Outbreak of Multidrug-Resistant *Salmonella* Infections Linked to Raw Turkey Products

Latest Outbreak Information

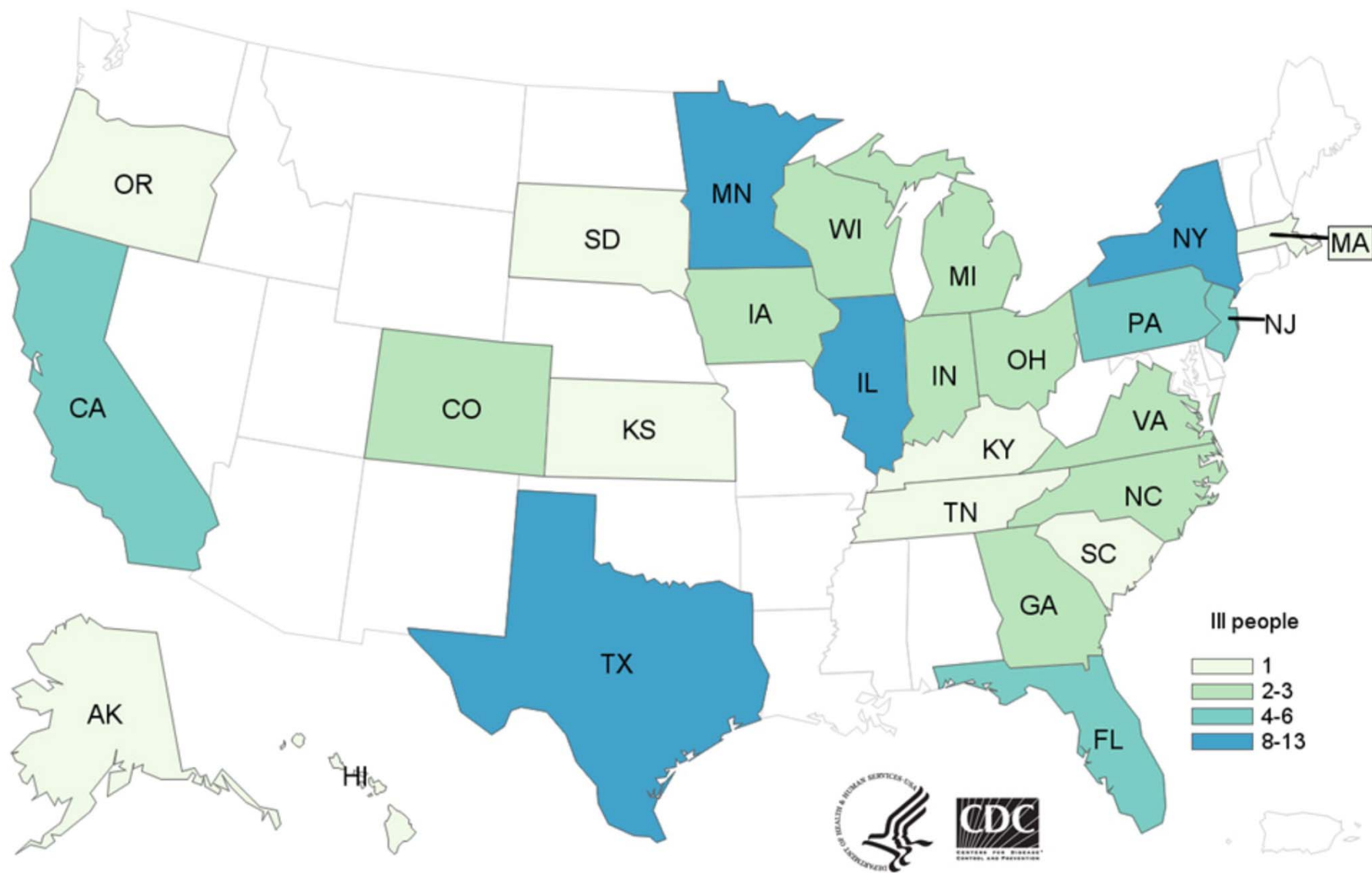


- Ninety people infected with the outbreak strain of *Salmonella* Reading have been reported from 26 states.
 - Forty people have been hospitalized, and no deaths have been reported.
- [Epidemiologic and laboratory evidence](#) indicates that raw turkey products from a variety of sources are contaminated with *Salmonella* Reading and are making people sick.
- In interviews, ill people report eating different types and brands of turkey products purchased from many different locations. Two ill people lived in a household where raw turkey pet food was fed to pets.
- The outbreak strain has been identified in samples taken from raw turkey pet food, raw turkey products, and live turkeys.
- A single, common supplier of raw turkey products or of live turkeys has not been identified.
- The outbreak strain of *Salmonella* Reading is present in live turkeys and in many types of raw turkey products, indicating it might be widespread in the turkey industry. CDC and USDA-FSIS have shared this information with representatives from the turkey industry and asked about steps that they may be taking to reduce *Salmonella* contamination.

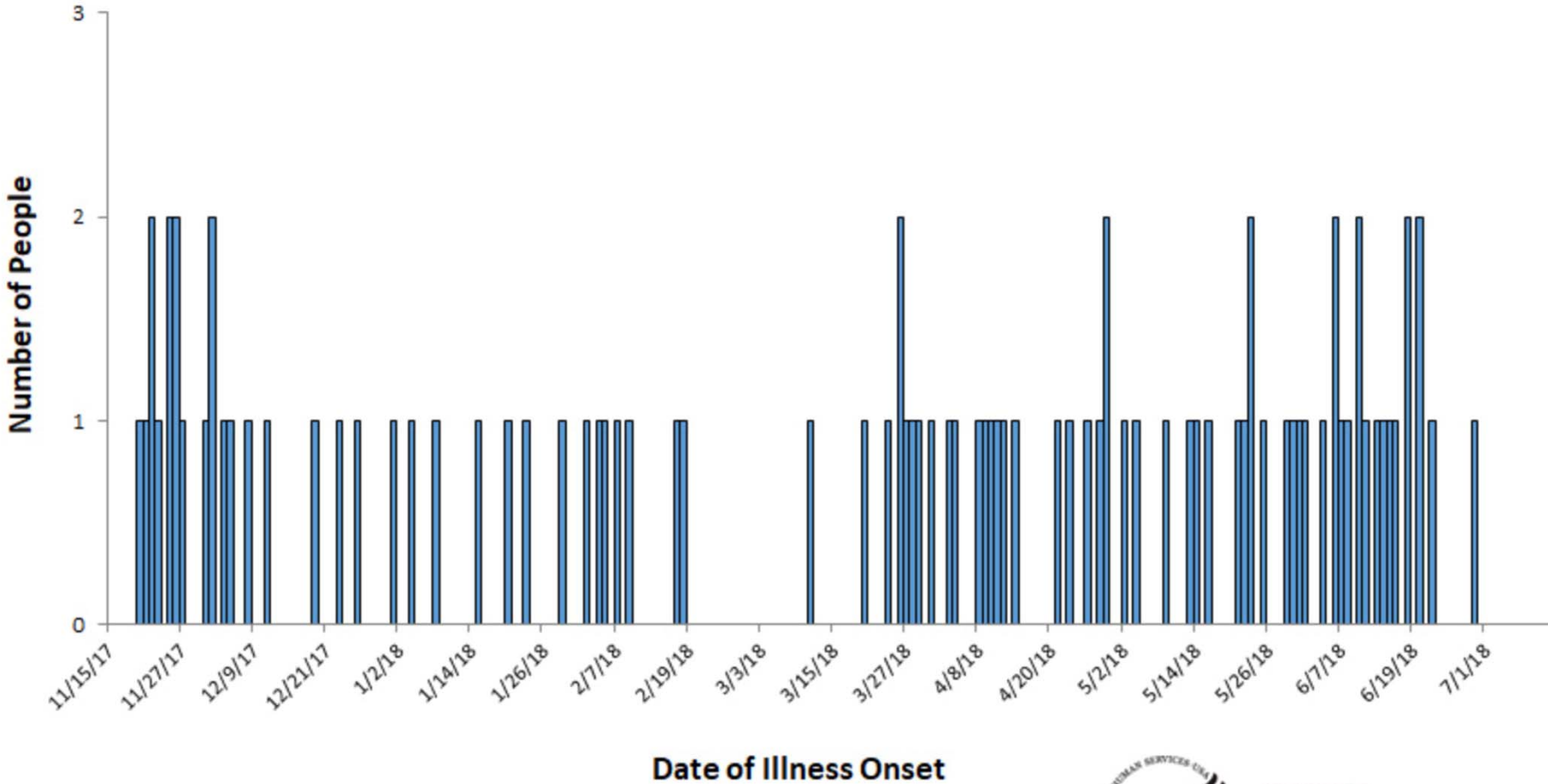
At A Glance

- [Reported Cases](#): 90
- [States](#): 26
- [Hospitalizations](#): 40
- [Deaths](#): 0

People infected with the outbreak strain of *Salmonella* Reading, by state of residence, as of July 11, 2018 (n=90)

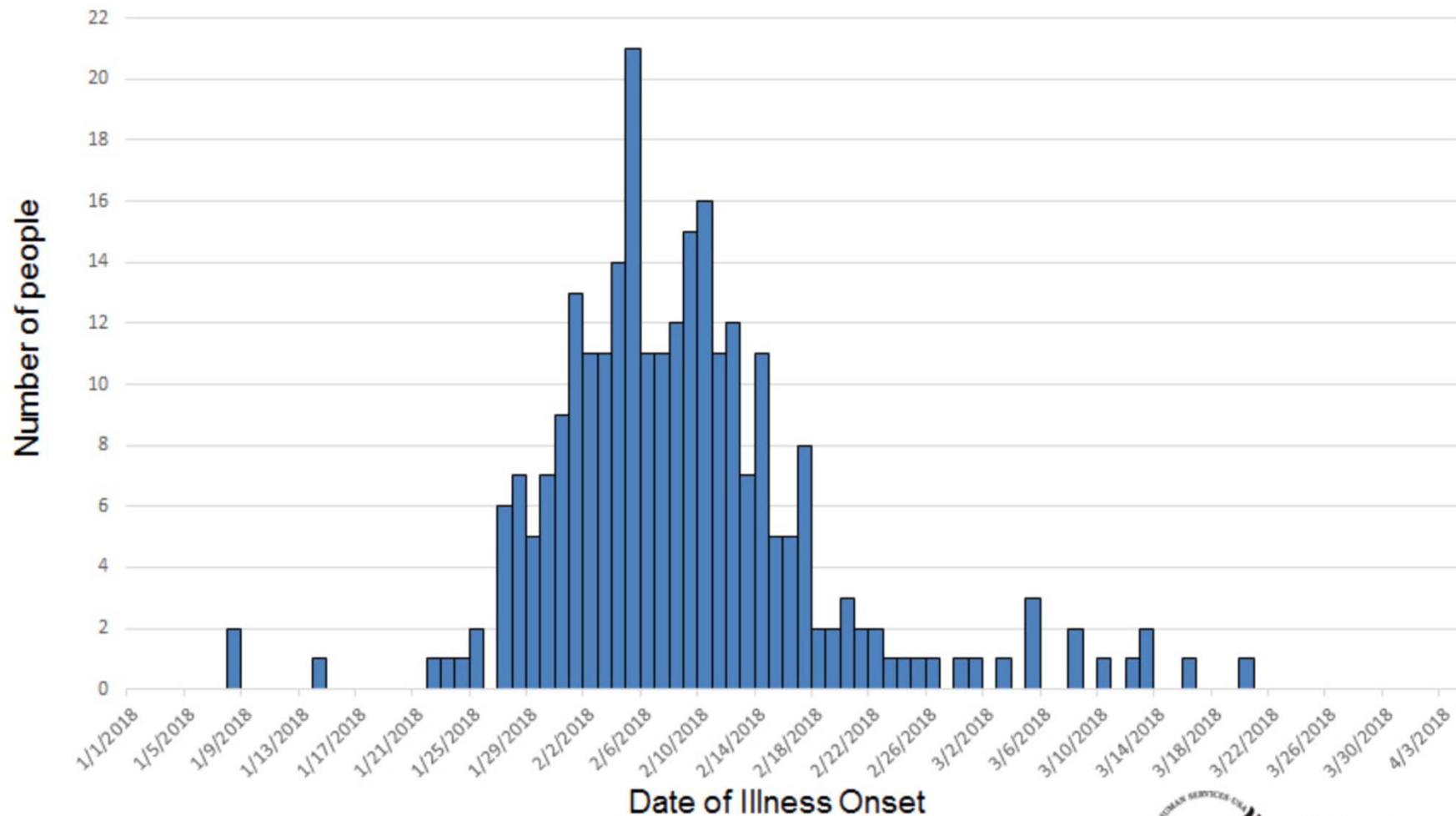


People infected with the outbreak strain of *Salmonella* Reading, by date of illness onset*

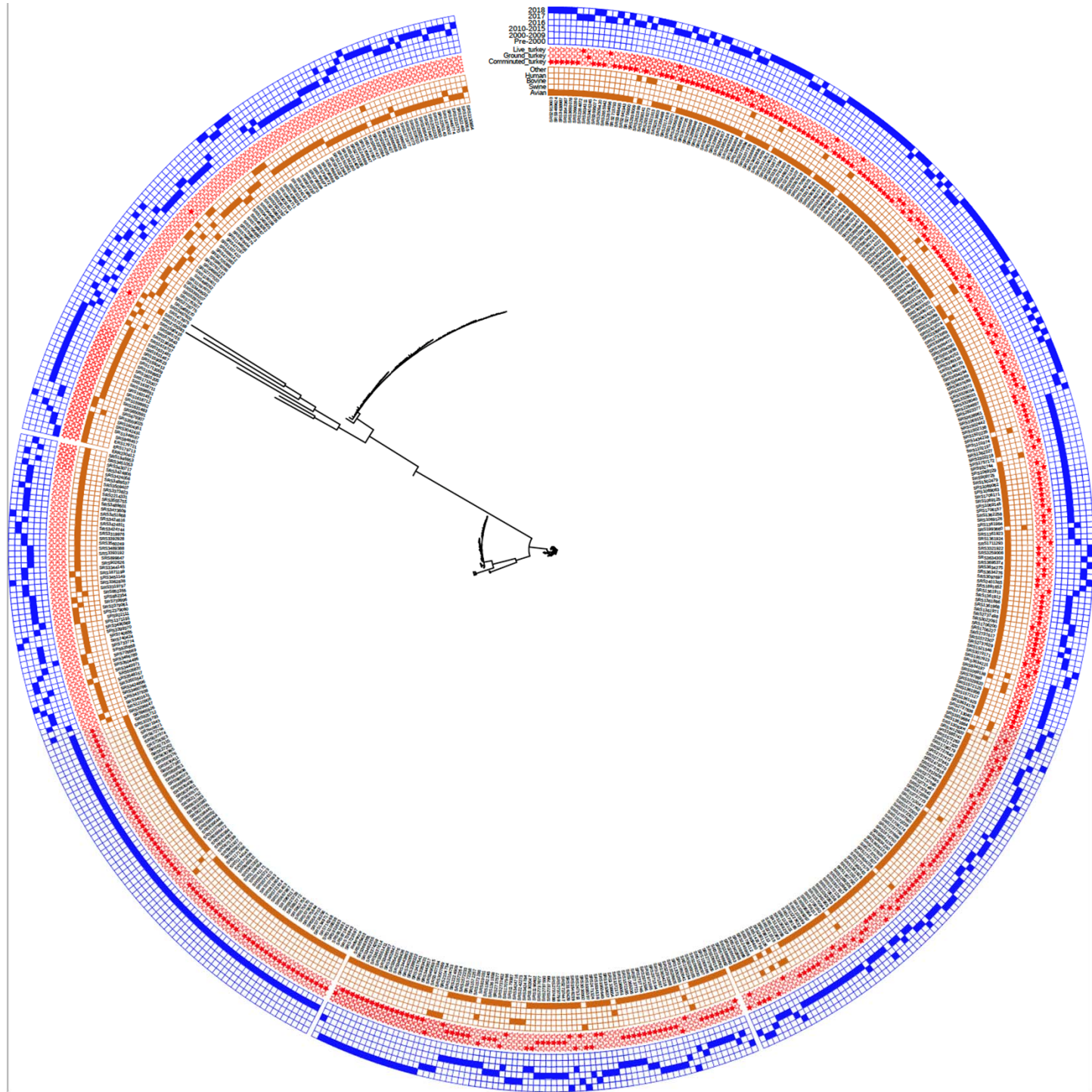


*n=90 for whom information was reported as of July 11, 2018. Some illness onset dates have been estimated from other reported information.

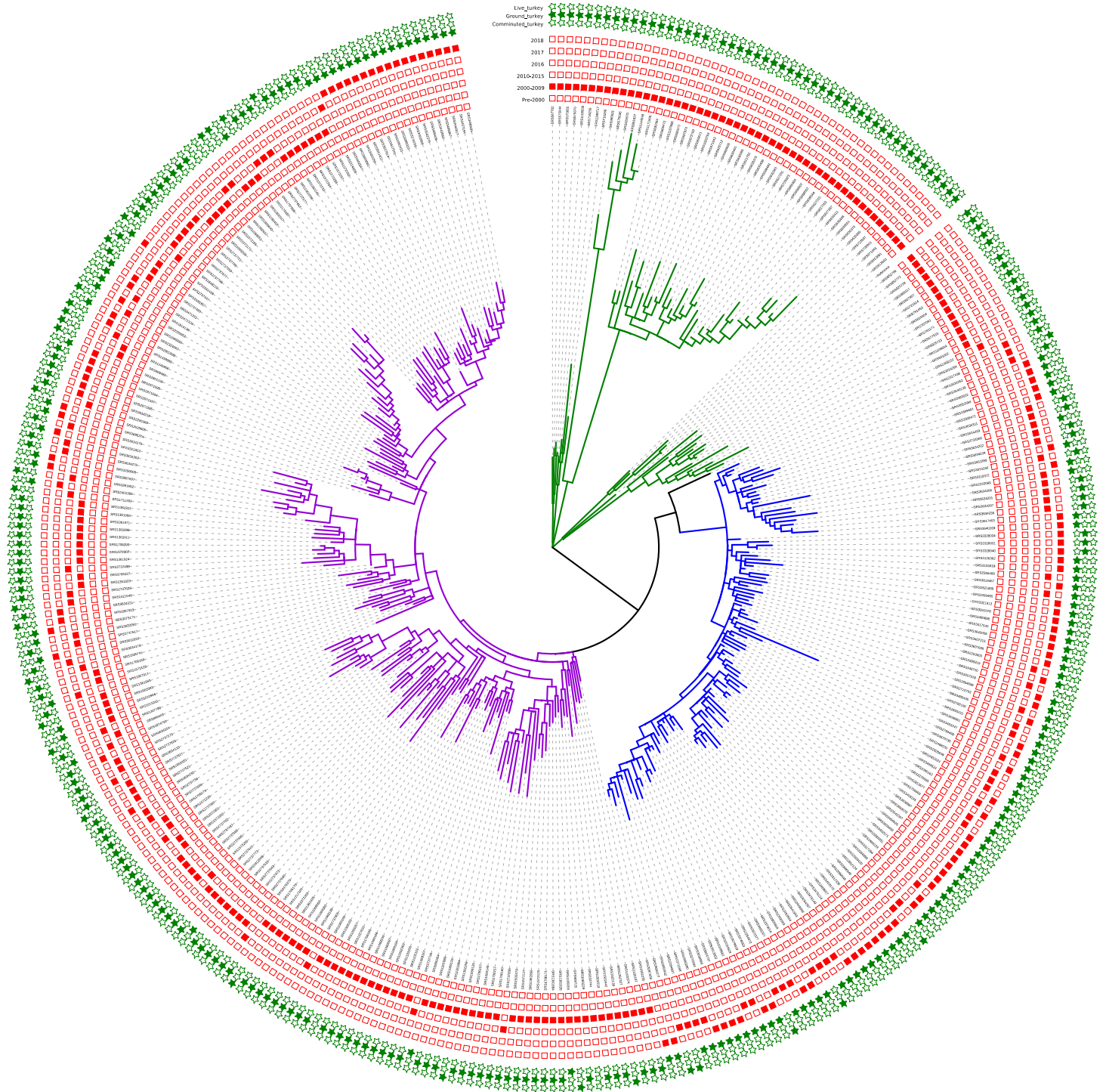
People infected with the outbreak strains of *Salmonella* Typhimurium, by date of illness onset*



*n=265 for whom information was reported as of April 4, 2018. Some illness onset dates have been estimated from other reported information.



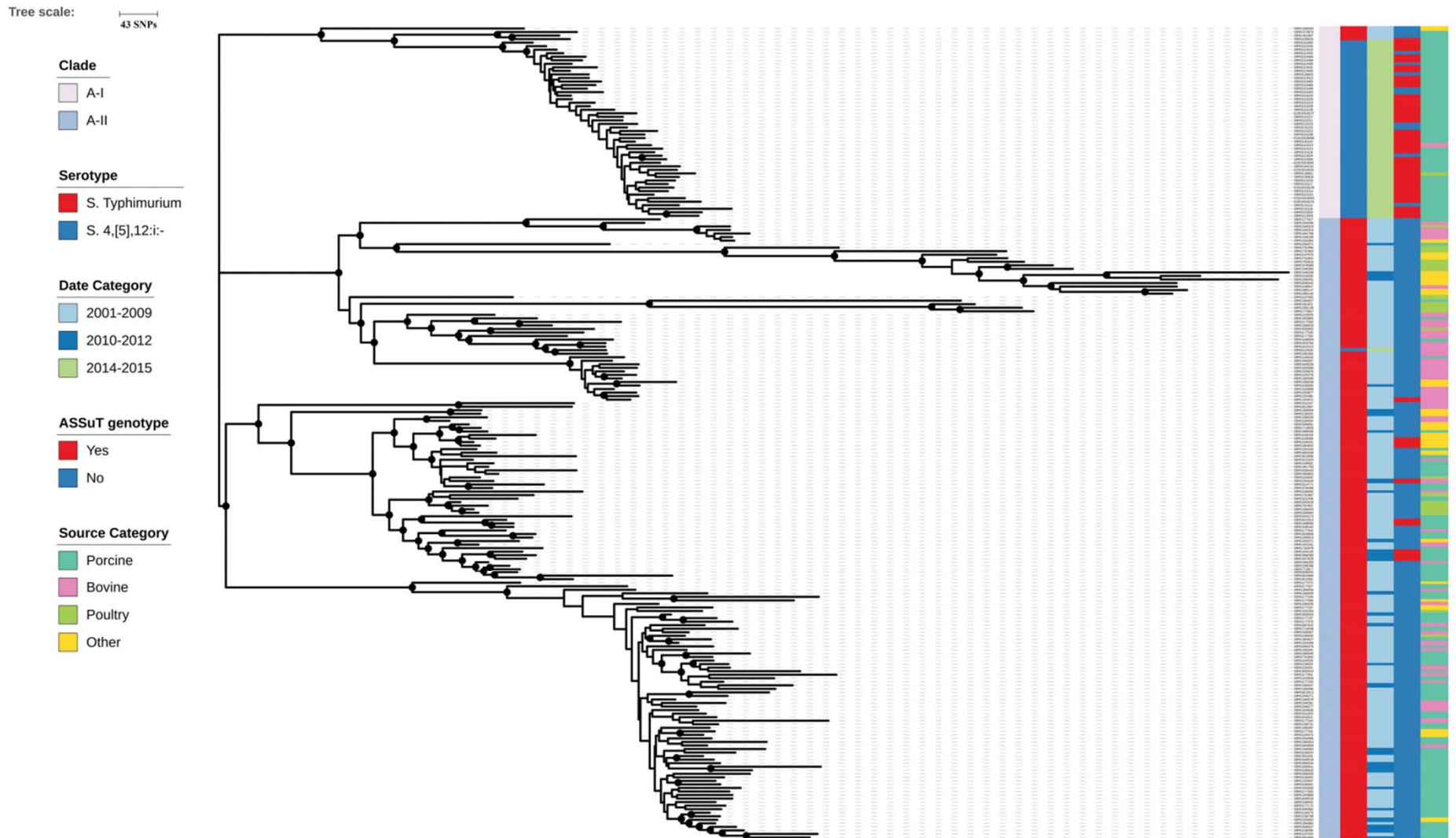
Tree scale: 100



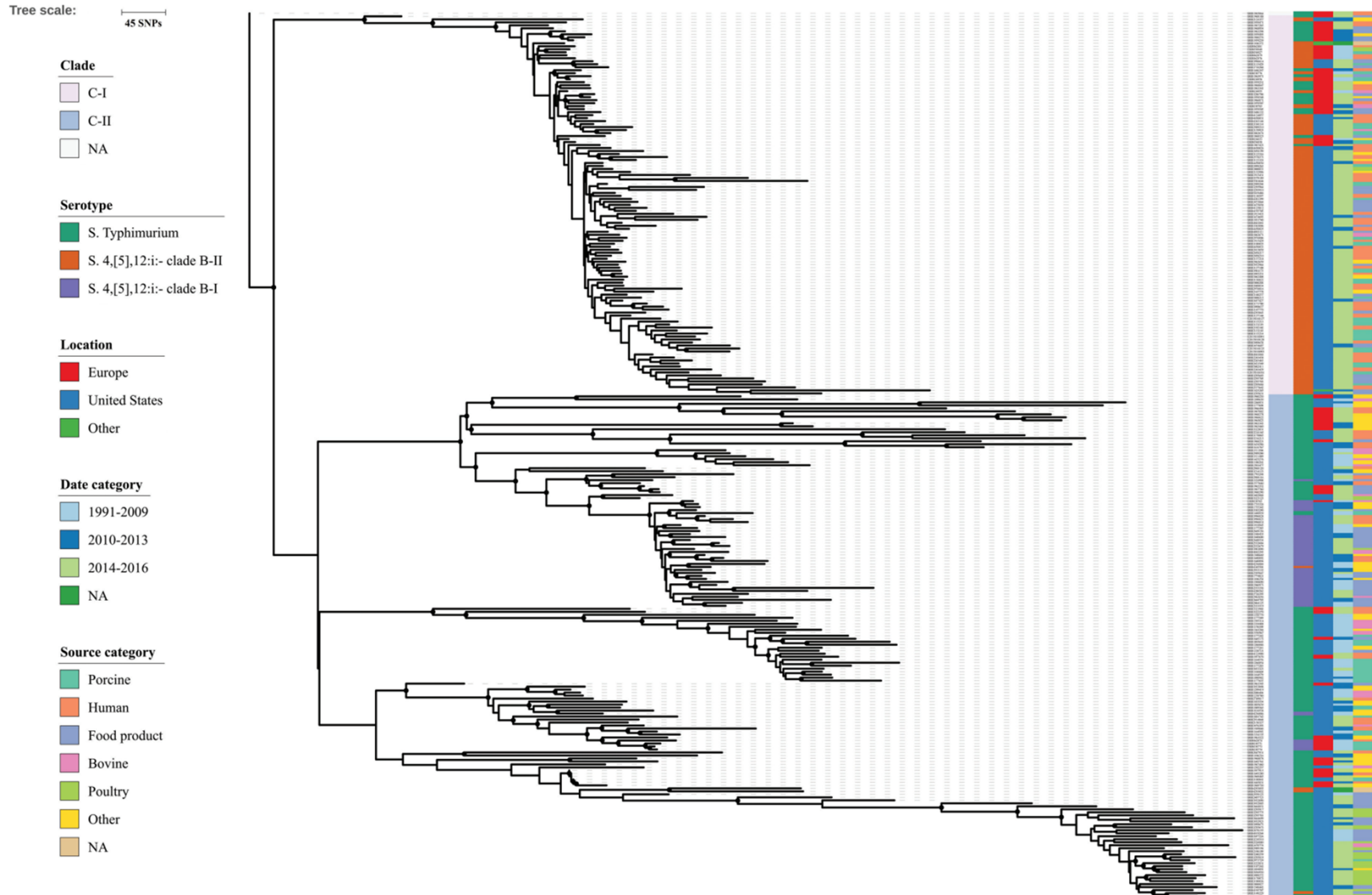
What I learned about the outbreak *Salmonella* Reading without any sampling of my own

- A new strain has emerged in the last two years
- The abundance of this strain is higher in turkey products than previous Reading strains
- It harbors a mobile DNA encoding drug resistance (ampicillin and tetracycline)
- Does not seem more virulent; however it has occupied a niche in turkeys

Salmonella 4,[5],12:i:- emergence in Europe and dissemination to USA



Salmonella 4,[5],12:i:- emergence in Europe and dissemination to USA



Take-home:

- *Salmonella* adapt and evolve to their surroundings
- Host adaptation versus host range are ongoing evolutionary forces
- Plasmids play a major role in adaptation
- Niche displacement occurs at both serovar and strain level